

# Evolutionary adaptations of plant AGC kinases: from light signaling to cell polarity regulation

Eike H. Rademacher and Remko Offringa\*

Molecular and Developmental Genetics, Institute Biology Leiden, Leiden University, Leiden, Netherlands

## Edited by:

Markus Geisler, University of Fribourg, Switzerland

## Reviewed by:

Christian Luschnig, University of Natural Resources and Life Sciences, Austria

Stephanie Robert, Umeå Plant Science Center, Sweden

## \*Correspondence:

Remko Offringa, Molecular and Developmental Genetics, Institute Biology Leiden, Leiden University, Sylviusweg 72, 2333BE Leiden, Netherlands.  
e-mail: [r.offringa@biology.leidenuniv.nl](mailto:r.offringa@biology.leidenuniv.nl)

Signaling and trafficking over membranes involves a plethora of transmembrane proteins that control the flow of compounds or relay specific signaling events. Next to external cues, internal stimuli can modify the activity or abundance of these proteins at the plasma membrane (PM). One such regulatory mechanism is protein phosphorylation by membrane-associated kinases, several of which are AGC kinases. The AGC kinase family is one of seven kinase families that are conserved in all eukaryotic genomes. In plants evolutionary adaptations introduced specific structural changes within the AGC kinases that most likely allow modulation of kinase activity by external stimuli (e.g., light). Starting from the well-defined structural basis common to all AGC kinases we review the current knowledge on the structure-function relationship in plant AGC kinases. Nine of the 39 *Arabidopsis* AGC kinases have now been shown to be involved in the regulation of auxin transport. In particular, AGC kinase-mediated phosphorylation of the auxin transporters ABCB1 and ABCB19 has been shown to regulate their activity, while auxin transporters of the PIN family are located to different positions at the PM depending on their phosphorylation status, which is a result of counteracting AGC kinase and PP6 phosphatase activities. We therefore focus on regulation of AGC kinase activity in this context. Identified structural adaptations of the involved AGC kinases may provide new insight into AGC kinase functionality and demonstrate their position as central hubs in the cellular network controlling plant development and growth.

**Keywords:** AGC kinase, evolution, kinase structure, *Arabidopsis thaliana*, PIN polarity, phototropic growth

## FUNCTIONAL DIVERSIFICATION OF PLANT AGC KINASES

Cellular responses to external or internal stimuli include the fast modification of the already present machinery of proteins, which might eventually trigger the activation of novel transcriptional programs. A commonly used modification in cellular signaling is protein phosphorylation, which is achieved by the addition of a phosphate group to the side chain of an amino acid. In eukaryotes, protein kinases typically catalyze the transfer of a gamma-phosphoryl group from adenosine triphosphate (ATP) to a serine, threonine, or tyrosine in their substrate proteins. Other amino acids such as histidine, arginine, aspartate, and lysine are uncommon phosphorylation targets in eukaryotes, but often utilized in prokaryotes.

Members of the protein kinase like superfamily have been found in genomes of archae, bacteria, and eukaryotes, highlighting the ancient origin of protein phosphorylation and its evolutionary conservation. With the arrival of multicellular eukaryotic organisms protein phosphorylation has been adopted to control a huge variety of cellular processes, and the involved kinases have evolved to fulfill new tasks, from triggering cell division to regulating membrane transport and cell polarity establishment. Eukaryotic protein kinases (ePKs) have been subdivided into 11 groups based on sequence similarity, evolutionary conservation, and known functions (Hanks and Hunter, 1995; Manning et al., 2002). Six of these subgroups (AGC, CAMK, CKI, CMGC, STE, PKL) are common

to 21 eukaryotic genomes covering fungi, animals, plants, apicomplexa, amoebozoia, red algae, and diatoms (Miranda-Saavedra and Barton, 2007).

AGC kinases are among the most well-studied kinases, and are named after the cyclic AMP dependent kinases (PKA), cGMP-dependent kinases, and the diacylglycerol-activated/phospholipid-dependent kinase PKC. In the model plant *Arabidopsis thaliana* the AGC group consists of 39 members (Table 1). The basal member of this group is the 3-phosphoinositide dependent protein kinase 1 (PDK1), which is highly conserved among eukaryotes. Due to its ability to activate other AGC kinases it is considered a master regulator of AGC kinase activity in mammalian cells (Mora et al., 2004). Furthermore, orthologs of the p70 ribosomal protein S6 kinase (S6K), the nuclear Dbf2-related (NDR) kinase subfamily, and the “AGC other” group of mammalian and yeast kinases can be found in *Arabidopsis*. However, the remaining 23 *Arabidopsis* AGC kinases seem to have no counterpart outside the plant kingdom, and are considered a separate subfamily (AGCVIII) in which groups AGC1 to 4 can be distinguished (Hanks and Hunter, 1995; Bögre et al., 2003; Galvan-Ampudia and Offringa, 2007).

Based on amino acid sequence homology these plant specific AGCVIII kinases are most closely related to animal PKA and PKC, which are involved in the regulation of cell polarity, -growth, and -division, and for which no direct homologs are

**Table 1 | AGC kinases of *Arabidopsis thaliana*.**

Locus	GenPept accession	Names	Full length	N-tail	Activation segment	C-tail	AGC kinase classification	
							1	2
At3g44610	NP_190047.2	AGC1–12	451	77	102	33	AGC1	AGCVIII
At4g26610	NP_194391.1	AGC1–2/D6PKL1	506	131	101	45		
At5g55910	NP_200402.1	AGC1–1/D6PK	498	117	105	47		
At5g47750	NP_199586.1	PK5/D6PKL2	586	199	104	54		
At3g27580	NP_189395.1	PK7/D6PKL3	578	190	102	57		
At2g44830	NP_850426.1	AGC1–3	765	371	102	63		
At5g40030	NP_198819.1	AGC1–4	499	122	98	51		
At1g16440	NP_173094.4	AGC1–6/RSH3	499	121	89	58		
At1g79250	NP_178045.2	AGC1–7	555	154	102	70		
At3g12690	NP_187875.1	AGC1–5	577	193	92	63		
At5g03640	NP_195984.1	AGC1–8	926	549	105	43		
At2g36350	NP_181176.1	AGC1–9	949	567	107	46		
At3g52890	NP_566973.2	KIPK	934	546	109	50		
At2g34650	NP_181012.1	PID	438	83	79	39	AGC3	
At2g26700	NP_180238.2	AGC3–4	526	95	137	55		
At1g53700	NP_175774.1	WAG1	476	101	73	71		
At3g14370	NP_188054.1	WAG2	480	96	75	79		
At3g45780	NP_190164.1	PHOT1	996	671	57	39	AGC4	
At5g58140	NP_851210.1	PHOT2	915	585	55	46		
At4g13000	NP_193036.1	AGC2–2	372	28	76	45	AGC2	
At3g25250	NP_189162.1	AGC2–1/OXI1	421	25	80	87		
At1g51170	NP_564584.1	AGC2–3/UCN	404	30	83	59		
At3g20830	NP_188719.1	AGC2–4/UCNL	408	29	85	62		
At4g14350	NP_193171.2	NDR-1	551	127	69	134	AGCVII	
At1g03920	NP_171888.1	NDR-2	569	145	68	129		
At3g23310	NP_188973.2	NDR-3	568	128	71	148		
At2g19400	NP_565453.1	NDR-4	527	113	77	114		
At2g20470	NP_179637.2	NDR-5	569	132	67	143		
At4g33080	NP_195034.2	NDR-6	519	102	77	112		
At1g30640	NP_174352.1	NDR-7	562	128	73	140		
At5g09890	NP_568221.1	NDR-8	515	110	72	112		
At5g62310	NP_201037.1	IRE	1168	762	59	120	AGC other	
At1g48490	NP_564529.4	IRE-3	1235	836	53	119		
At1g45160	NP_175130.2	IRE-4	1042	678	55	82		
At3g17850	NP_188412.2	IRE-H1	1296	890	59	120		
At3g08720	NP_187485.1	S6K1	465	142	27	70	AGCVI	
At3g08730	NP_187484.1	S6K2	471	148	27	71		
At5g04510	NP_568138.1	PDK1–1	491	50	38	174	PDK1	
At3g10540	NP_187665.2	PDK1–2	486	51	38	168		

The table represents all 39 AGC kinases found in *Arabidopsis*. The first three columns give the TAIR locus and GenPept accession numbers for each kinase as well as some of the names assigned to the genes, respectively. The fourth column represents the full length of the respective amino acid sequence and is followed by the length of the N-tail (counted until the second glycine of the conserved GxGxxG motive in subdomain I), the length of the activation segment (DFD/GLS to APE), and the length of the C-tail (counted from the tryptophan of the FxxxW motive in subdomain XI). The last two columns represent the classification of *Arabidopsis* AGC kinases 1) as proposed by Galvan-Ampudia and Offringa (2007), 2) based on earlier classifications (Hanks and Hunter, 1995; Bögre et al., 2003).

found in plants. Plant AGCVIII kinases have evolved into regulators of a variety of developmental processes and stress responses. In unicellular green algae, such as *Chlorella variabilis*, *Ostreococcus tauri*, and *Chlamydomonas reinhardtii*, orthologs of the *Arabidopsis* PHOTOTROPIN 2 (PHOT2) are the only AGCVIII

kinases that can be identified, indicating that these are the direct descendants of the first ancestral AGCVIII protein kinase (Onodera et al., 2005; Derelle et al., 2006; Galvan-Ampudia and Offringa, 2007; Blanc et al., 2010). This implies that the ancestral plant AGCVIII kinases acquired an N-terminal regulatory

photoreceptor domain that was lost in later descendants. Interestingly, phot2 in land plants is involved in responses to high light intensity, such as light avoidance of chloroplasts and actin-dependent positioning of the nucleus (Iwabuchi et al., 2010), which are typically functions that are required in unicellular plant life. Later in evolution, a second phototropin (PHOT1) co-occurred with the first appearance of seed plants (Galvan-Ampudia and Offringa, 2007). Phot1 mediates responses to low intensity blue light (BL) and exclusively mediates BL-dependent inhibition of hypocotyl elongation of germinating seeds reaching the substrate surface (Folta and Spalding, 2001). Combined these two AGC4 kinases enable seed plants to optimize their photosynthetic efficiency and subsequent growth in response to changing light conditions.

Closely related but distinct from the other AGCVIII kinases are the four AGC2 kinases OXIDATIVE SIGNAL-INDUCIBLE1 (OXI1/AGC2-1), AGC2-2, UNICORN (UCN/AGC2-3), and UNICORN-LIKE (UCNL/AGC2-4). OXI1 and AGC2-2 have been shown to be involved in root growth (Anthony et al., 2004; Camehl et al., 2011), oxidative stress signaling (Rentel et al., 2004), and plant defense responses (Petersen et al., 2009). A recent report indicated that UCN and UCNL regulate cell growth and division in integuments, the embryo proper, cotyledons, and floral organs (Enugutti et al., 2012).

The remaining 17 AGCVIII kinases can be subdivided in two groups, AGC1 and AGC3 (Galvan-Ampudia and Offringa, 2007). Of the AGC3 kinases PINOID (PID), WAG1, and WAG2 have been shown to regulate the polarity of auxin transport by phosphorylating the large central hydrophilic loop (HL) of PIN-FORMED (PIN) auxin efflux carriers (Michniewicz et al., 2007; Dhonukshe et al., 2010; Huang et al., 2010). The role of the fourth kinase AGC3-4/PID2 is still ambiguous, as based on the current data it is still unclear whether this kinase acts redundantly with the other three AGC3 kinases during embryo development (Cheng et al., 2008; Dhonukshe et al., 2010). Also four of the AGC1 kinases (D6 PROTEIN KINASE (D6PK)/AGC1-1, D6 PROTEIN KINASE LIKE 1 (D6PKL1)/AGC1-2, D6PKL2/PK5, D6PKL3/PK7) have been implicated to have a regulatory role in polar auxin transport. Although they can phosphorylate the PIN-HL *in vitro*, their exact function is still unclear (Zourelidou et al., 2009). AGC1-5 and AGC1-7 have been implied in polarized pollen tube growth (Zhang et al., 2009). To our knowledge the role of the other 7 *Arabidopsis* AGC1 kinases has not been determined so far, except that KCB INTERACTING PROTEIN KINASE (KIPK) was found to interact with a kinesin-like protein (Day et al., 2000). In tomato, the AGC1 kinase AvrPto-DEPENDENT Pto-INTERACTING PROTEIN 3 (Adi3) has been implied as possible ortholog of mammalian Akt/PKB, as both proteins act as negative regulator of programmed cell death (PCD; Devarenne et al., 2006). Based on homology the putative negative regulator of PCD in *Arabidopsis* would be AGC1-3.

## STRUCTURAL FEATURES OF PLANT AGC KINASES

Misregulation of AGC kinase activity in mammalian systems causes the development of severe diseases including cancer and diabetes. Naturally, this has attracted intense research and has led to the elucidation of various crystal structures of human and

mammalian AGC kinases as well as the fine mapping of relevant subdomain functions (Taylor and Kornev, 2011).

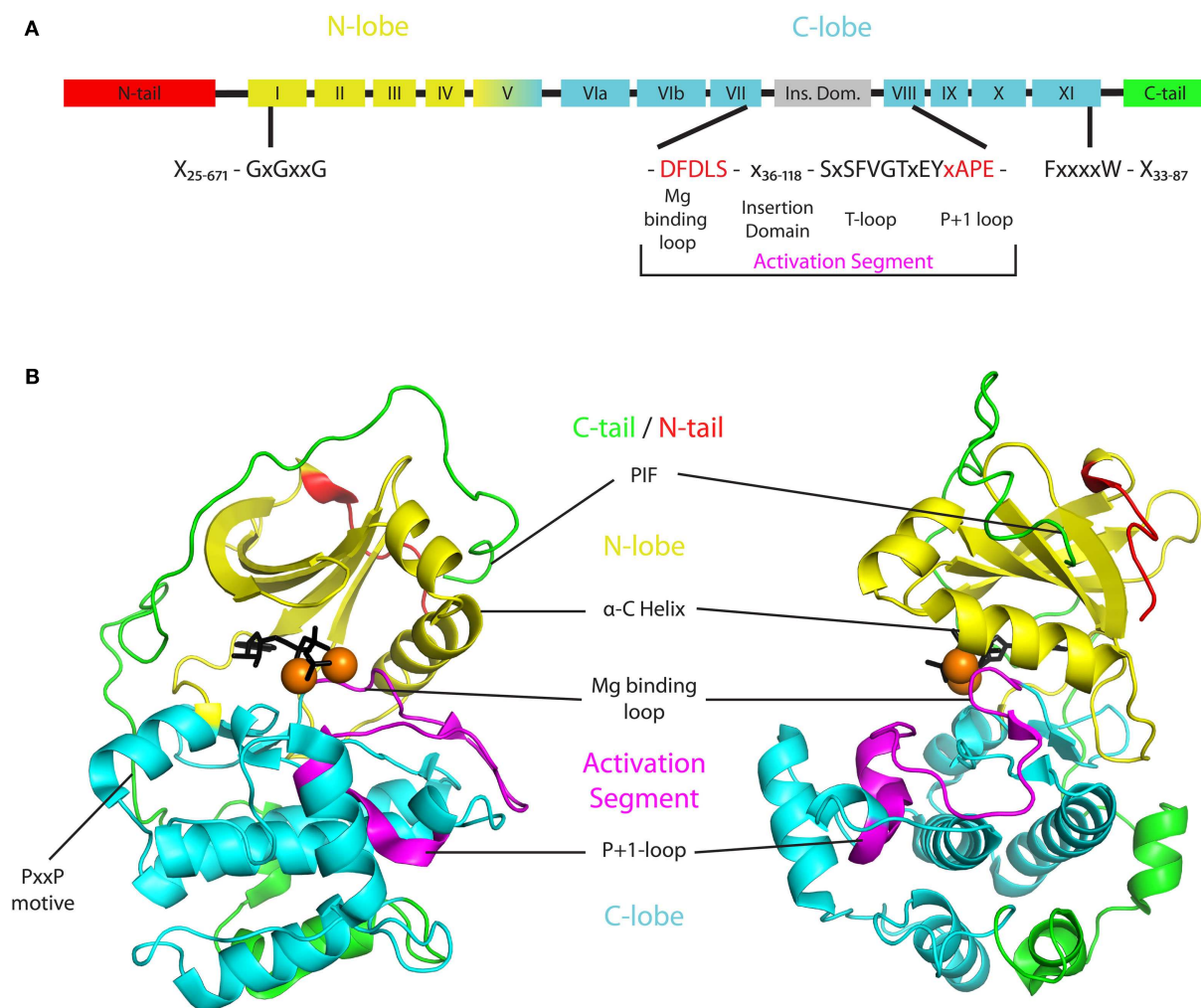
Of the *Arabidopsis* AGC kinases only phot1 and phot2 have been studied in structural detail, with particular emphasis on the photosensory domains. A recently published structure of a phot2 fragment comprising the kinase domain and one of the photosensory domains remains the only available plant AGC kinase structure to date (Takayama et al., 2011). The structure of phot2 underlines that evolutionary conservation of structural elements can be found within the group of AGC kinases. Hence, it should be possible to draw conclusions on the structure and regulation of plant AGC kinases based on what is known from well-researched animal AGC kinases.

## THE CATALYTICAL CORE OF AGC KINASES

Like all protein kinases, plant AGC kinases contain the universal catalytic core that is built up of 12 conserved subdomains (Figure 1A; Hanks and Hunter, 1995). This catalytic core binds ATP together with  $Mg^{2+}$  and catalyzes the transfer of a phosphate group onto a substrate protein/amino acid. It is formed by two lobes that are interconnected by a linker domain (Figure 1). The smaller N-terminal lobe (subdomains I–IV) is dominated by five  $\beta$ -strands but also features a prominent conserved  $\alpha$ -helix, called  $\alpha$ -C helix (subdomain III). The larger and more rigid C-terminal lobe on the other hand is mainly formed by  $\alpha$ -helices (subdomains VIa–XI). The flexible linker domain between the two lobes (subdomain V) functions as a hinge and allows for rotation of the lobes and subsequent opening or closure of the ATP binding pocket located in the cleft between them (Figure 1B). Correct positioning of a highly conserved glycine-rich loop (GxGxxG, P-loop) in subdomain I of the N-lobe and an invariant aspartate in subdomain VII of the C-lobe is mandatory for ATP binding in the catalytic cleft. Furthermore subdomains VIB and VIII within the C-lobe form the binding sites for protein substrates (Hanks and Hunter, 1995; Nolen et al., 2004; Taylor and Kornev, 2011). Accordingly, binding of ATP and substrates is regulated by conformational changes in the catalytic core. These changes are evoked by phosphorylation of accessory domains protruding from the core at the N- or C-terminus (called N- or C-terminal tail), and of the activation segment that comprises the residues between two conserved tripeptides (DFG and APE) found in respectively subdomains VII and VIII of the C-lobe.

## REGULATORY DOMAINS IN THE N-TERMINAL TAIL

The N-terminal tail (N-tail) is highly variable between AGC kinases and extends over the N-lobe. While the N-tail of the PDK1s and AGC2 kinases is relatively short (25–50 amino acids) it is relatively large in several other AGC kinases (up to 890 amino acids in IREH-1; Table 1). This makes it likely that the N-tail is a site for post-translational regulation of kinase activity, either by different modifications such as myristoylation, deamidation, and phosphorylation, or by binding of regulatory cofactors. Even though such modifications and interactions have been described for mammalian PKA and Akt/PKB (Currie et al., 1999; Tholey et al., 2001; Sastri et al., 2005; Bastidas et al., 2012), functional studies on plant AGC N-tails are largely lacking. Only the N-tails of PHOT1 and PHOT2 have been analyzed in greater detail and



**FIGURE 1 | Basic structure of AGCVIII kinases. (A)** Two dimensional structure indicating the different protein domains of AGCVIII kinases. The N- and C-lobe (yellow or cyan) of the catalytic core are supplemented by regulatory domains in the N-tail (red), activation segment (magenta), and C-tail (green). These three domains are highly variable and their minimal and maximal lengths in AGCVIII kinases are denoted [N-tail 25–671 amino acids (AA), activation segment 55–137 AA, C-tail 33–87 AA]. Importantly, the magnesium binding loop (DFDLS) and the activation loop (SxSFVGTxEY) are separated by a 36–118 AA long insertion domain that is found in plant AGC kinases only. **(B)** Exemplary three dimensional structure of an AGC kinase with the same color coding as in the two dimensional structure. The depicted structure is based on a ternary crystal structure of mouse PKA [PDB-ID: 4DG2 (Bastidas et al., 2012)]. The beta-sheet rich N-lobe (yellow) is connected to the alpha-helix rich C-lobe (cyan) by a linker formed by subdomain V. Together they form the catalytic

core of AGC kinases and enable ATP (black), magnesium (orange spheres), and substrate (not shown) binding in the catalytic cleft between the lobes. The activity of AGC kinases is regulated by post-translational modification of residues in, or binding of interactors to the N-tail (red), activation segment (magenta), or C-tail (green). The N-tail depicted here represents the anchor of a, in some cases much bigger, regulatory domain that extends from the core structure. Also the activation segment and the C-tail are highly diverse. The activation segment starts with a magnesium binding loop (DFDLS, black) and ends with the  $P + 1$  loop (APE, purple) that is involved in substrate binding. Different from the depicted situation in PKA, a quite variable insertion domain of up to 118 AA can be found within the activation segment of plant AGC kinases. Likewise, the C-tail of most plant AGC kinases contains conserved elements, such as the PxxP motif or the PIF at its C-terminus (green), but also shows great diversity in composition and size.

photosensory domains as well as various phosphorylation sites have been mapped (for review see (Christie, 2007)).

#### THE MULTIFUNCTIONAL ACTIVATION SEGMENT IS EXTENDED IN PLANT AGC KINASES

The activation segment of protein kinases generally contains a magnesium binding loop (DFG), a T-loop (also called activation loop), and a  $P + 1$  loop (xAPE, Figure 1A; Nolen et al., 2004;

Taylor and Kornev, 2011). The latter forms a critical interaction point between substrate and kinase, while the aspartate in the DFG motif of the magnesium binding loop is responsible for chelating one of the  $Mg^{2+}$ -ions that orient the ATP for phospho-transfer. The following phenylalanine contributes to the formation of the kinase – substrate contact surface by interacting with residues of the N-lobe (Nolen et al., 2004). While these two residues are invariant in all AGC kinases, the glycine in the DFG motif has been



exchanged for an aspartate in plant AGCVIII kinases. This amino acid change appears to reduce kinase activity *in vitro* (Christensen et al., 2000).

In AGC kinases, phosphorylation of a serine or threonine residue in the T-loop ([S/T]<sup>P</sup>xxGTx[D/E]Y) is necessary to activate the kinase (Chan et al., 1999; Bögre et al., 2003). Phosphorylation of the T-loop results in a conformational change involving the  $\alpha$ -C helix (**Figure 1B**) that stabilizes the kinase in an open structure allowing substrate binding (Huse and Kuriyan, 2002). Vice versa, when the T-loop is in an inactive conformation a preceding beta-sheet becomes disordered and can no longer anchor the activation segment in the catalytic cleft. Subsequently, binding of Mg<sup>2+</sup> and substrate peptides is inhibited (Nolen et al., 2004).

Interestingly, in the plant specific AGCVIII kinases a larger insertion of 36–118 amino acids in the activation segment, also referred to as the T-loop extension (Bögre et al., 2003), VII–VIII insertion (Galvan-Ampudia and Offringa, 2007), or insertion domain (Zegzouti et al., 2006b), separates the Mg<sup>2+</sup> binding site (DFDLs) and the T-loop (SxS<sup>P</sup>FVGTxEY; **Figure 1A**). This insertion domain is typical for plant AGC kinases, and for some of the AGC kinases it was found to determine their subcellular localization. The insertion domain of *Arabidopsis* PID kinase has been shown to direct its plasma membrane (PM) association (Zegzouti et al., 2006b), and for the tomato AGC kinase Adi3 it contains a nuclear localization sequence (NLS) (Ek-Ramos et al., 2010). For other kinases the function of the insertion domain is unclear, but it is likely that it provides an additional structural feature to the kinase that allows further regulation of its activity or its subcellular localization.

In most of the AGCVIII kinases the insertion domain contains a conserved AEP motif directly linked to the T-loop. The function of this AEP motif is not clear, but the fact that it can also be found in PHOT2 orthologs in unicellular algae, confirms that AGCVIII kinases are direct descendants from an ancestral phototropin. Its absence in the AGC2 kinases suggests that this group branched off at a certain point in evolution from the other AGCVIII kinases by losing this feature. It will be interesting to see if the AEP motif is a hallmark for the functional divergence between these two groups of plant AGC kinases.

### THE C-TERMINAL TAIL PROVIDES PROTEIN-PROTEIN INTERACTION SITES

Other than the N-tail and the activation segment that both emerge from the catalytic core and form relatively independent subunits, the C-terminal tail (C-tail) stretches from the C-lobe around the kinase core and eventually folds back on to the N-lobe (**Figure 1B**). Three conserved segments have been identified within the C-tail that are shared by most AGC kinases. These are the C-lobe tether, the active-site tether, and the N-lobe tether (Kannan et al., 2007). All three have been implicated to be involved in mediating the opening and closing of kinases by interacting with conserved residues in the catalytic core. Regulatory motifs in these segments might serve as interaction sites for other factors. For example, a conserved PxxP motif in the C-lobe tether of mammalian Akt/PKB has been shown to serve as a binding site for its activating tyrosine kinase Src (Jiang and Qiu, 2003). Furthermore, a hydrophobic motive (FxxF) at the C-terminus of the N-lobe tether was found to

mediate interaction with PDK1 and was therefore named PDK1 interacting fragment (PIF, **Figure 1B**; Etchebehere et al., 1997; Biondi et al., 2000; Frodin et al., 2002). In some AGC kinases this motif is extended by a phosphorylation site (FxxF[S/T]Y), the phosphorylation of which enhances PDK1 binding and hence allows for an additional level of regulation (reviewed in Biondi, 2004). Next to mediating PDK1 interaction, *in vitro* reconstitution assays suggest that the PIF acts synergistically with T-loop phosphorylation in stimulating kinase activity. Potentially it does so by stabilizing the  $\alpha$ C helix in an active conformation that allows for optimal transfer of the ATP phosphate group to the substrate (Frodin et al., 2002).

### REGULATION OF PLANT AGC KINASES ACTIVITY

The activity of protein kinases is regulated by posttranslational modification and by interacting proteins or macromolecules, both of which have an effect on their enzymatic activity and/or on their subcellular localization. The latter in turn determines the proximity to and thus the chance to phosphorylate their downstream targets. Most of these regulatory aspects involve the N- or C-terminal tail and the activation segment of the protein kinase. For the plant AGCVIII kinases, considerable research has been done on the BL receptors phot1 and phot2, and events leading to their activation are relatively well understood. Due to their role in regulating polar auxin transport, also PID, and two other AGC3 protein kinases (WAG1 and WAG2) have been analyzed in more detail. Compared to the phototropins, however, crystal structures and a detailed investigation of regulatory domains of the AGC3 protein kinases are missing. Below we will therefore summarize the current knowledge, and speculate on possible regulatory pathways with a focus on the AGC3 kinases and their role in PIN trafficking.

### PDK1 AS REGULATOR OF AGC KINASE ACTIVITY

As indicated in Section “The Multifunctional Activation Segment is Extended in Plant AGC Kinases,” AGC kinases require phosphorylation of the T-loop for their activation. In mammalian systems several AGC kinases are phosphorylated by PDK1 that acts as a master regulator of AGC kinase activity. Other AGC kinases, including PDK1, are able to auto-activate through cis- or trans-autophosphorylation.

For PDK1, however, cis- and trans-autophosphorylation of its T-loop is not sufficient for activation, since it is initially auto-inhibited by its pleckstrin homology (PH) domain. This phosphoinositide binding domain bends over the catalytic cleft and blocks docking of protein substrates (Casamayor et al., 1999; Wick et al., 2003; Gao and Harris, 2006). In mammalian cells such “primed” but inactive PDK1 molecules reside in the cytoplasm as homo-dimers, interacting via the PH domain, ATP loaded, and with the T-loop phosphorylated. Binding of the PH domain to 3-phosphoinositides in the PM leads to dissociation into monomers and stimulates trans-autophosphorylation of a threonine close to the PH domain. This may stabilize an open conformation of the activated PDK1 monomers, which are the active PM-associated forms that interact with and phosphorylate target AGC kinases, such as Akt/PKB (Wick et al., 2003; Masters et al., 2010).

PDK1 interacts with the hydrophobic motif in the PIF domain of its substrate kinases (**Figure 1B**) through a PIF binding pocket

present in its N-lobe (Biondi et al., 2000, 2001). Human AGC kinases often have a serine or threonine residue directly following their PIF domain (FxxF/Y[S/T]<sup>P</sup>F/Y) that needs to be phosphorylated before interaction with PDK1 can occur. In the mammalian AGC kinases PKC $\zeta$  and PKC related kinase 2 (PRK2) an aspartic acid and glutamic acid residue are respectively present at the serine/threonine position, and these residues are required for binding of and activation by PDK1, probably because they mimic the negative charge of a phosphorylated serine/threonine (Balendran et al., 2000).

Two *Arabidopsis* homologs of mammalian PDK1 (PDK1–1 and PDK1–2) have been identified. Based on the presence of a PIF domain in several plant AGC kinases, as well as on protein–protein interaction studies and *in vitro* activity assays, *Arabidopsis* PDK1 potentially is involved in T-loop phosphorylation and activation of at least 16 AGC kinases (Bögre et al., 2003; Anthony et al., 2004; Zegzouti et al., 2006a,b). Interestingly, most of these AGC kinases terminate in the tetrameric hydrophobic motif (FxxF) of the PIF domain and lack an adjacent serine/threonine residue.

Exceptions to this are the PIFs of AGC1–12 and S6K1 and 2. In the case of AGC1–12 the FxxF motif is hidden approximately 90 amino acids upstream of the C-terminus. In S6K1 and 2 the FxxF motif is found 15 amino acids upstream of the C-terminus. Recently it has been shown that the hydrophobic motif of the S6Ks is expanded to a TARGET OF RAPAMYCIN (TOR) phosphorylation motif of the form FxxFT<sup>P</sup>YVxP, and that this motif is phosphorylated by TOR kinase in *Arabidopsis* (Xiong and Sheen, 2012). Analogous to mammalian systems *Arabidopsis* TOR has been demonstrated to interact with S6K1 via REGULATORY-ASSOCIATED PROTEIN OF TOR (RAPTOR). Osmotic stress leads to removal of RAPTOR and the subsequent loss of PIF domain phosphorylation by TOR, which inhibits PDK1 dependent activation of S6K1 (Mahfouz et al., 2006).

Recently, doubts were raised, based on different observations, as to whether PDK1 is a master regulator of the plant specific AGCVIII kinases (Zhang and McCormick, 2009). A similar discussion is ongoing for the PKA isoforms, the closest animal homologs of the AGCVIII kinases. *E. coli* expressed PKA has been reported to auto-phosphorylate *in vitro* (Yonemoto et al., 1997), but other reports suggest that activation of PKA requires the activity of PDK1 (Cheng et al., 1998; Nirula et al., 2006). Interestingly, like the plant AGCVIII kinases, PKA also has a tetrameric FSEF motif at its C-terminus. It is thus tempting to speculate that kinases with the tetrameric C-terminal hydrophobic motif are less or even not dependent on PDK1 for their activation. In line with this hypothesis, PID:GUS or PID:VENUS translational fusions in which the FDYF motif was excluded were able to complement the *pid* loss-of-function mutant (Benjamins et al., 2001; Michniewicz et al., 2007). Although it has been shown *in vitro* that the PIF domain is essential for a functional interaction with PDK1 (Zegzouti et al., 2006a), the relevance of this domain and of PDK1 for the *in planta* function of PID still needs confirmation. PID can activate itself through intramolecular autophosphorylation, which may be sufficient *in planta* under optimal growth conditions. The *in vitro* observed 6.5-fold increase in PID phosphorylation in the presence of PDK1 may have a function in stressed plants (Zegzouti et al., 2006a). Also the close PID homologs WAG1 and WAG2 are

able to auto-phosphorylate, despite the fact that they do not have a C-terminal PIF domain and are not hyperactivated by PDK1 *in vitro*. Unexpectedly, they do interact with PDK1 in *in vitro* pull downs, and addition of the FDYF motif to the C-terminus of WAG1 enhances its autophosphorylation, whereas the presence of PDK1 reduces its activity (Zegzouti et al., 2006b). In line with this observation, the autophosphorylating AGC1 kinase KIPK has a FxxF motif and interacts with PDK1 but is not hyperactivated by PDK1 *in vitro* (Zegzouti et al., 2006b). All these data suggest that the interaction with PDK1 is not necessarily mediated by the FxxF motif, and that other residues may be important.

The *Arabidopsis* phototropins PHOT1 and PHOT2 lack a PIF domain (Bögre et al., 2003) and like WAG1 and WAG2 seem to act independently of PDK1. Both have been shown to auto-phosphorylate their N-terminal photosensory domain and their T-loop in response to BL (Salomon et al., 2003; Inoue et al., 2008). This action is controlled by two accessory light, oxygen, or voltage sensing (LOV) domains at the N-terminus of the kinases, of which LOV2 under dark conditions represses kinase activity by folding over the catalytic core. The LOV domains bind flavinmononucleotides (FMNs) as chromophores to sense BL, which induces the formation of a covalent adduct between FMN and a cysteine residue in each LOV domain (Salomon et al., 2000; Swartz et al., 2001). This results in dissociation of the LOV2 domain from the kinase domain, and leads to receptor dimerization and subsequent trans-phosphorylation as shown for phot1 (Kaiserli et al., 2009).

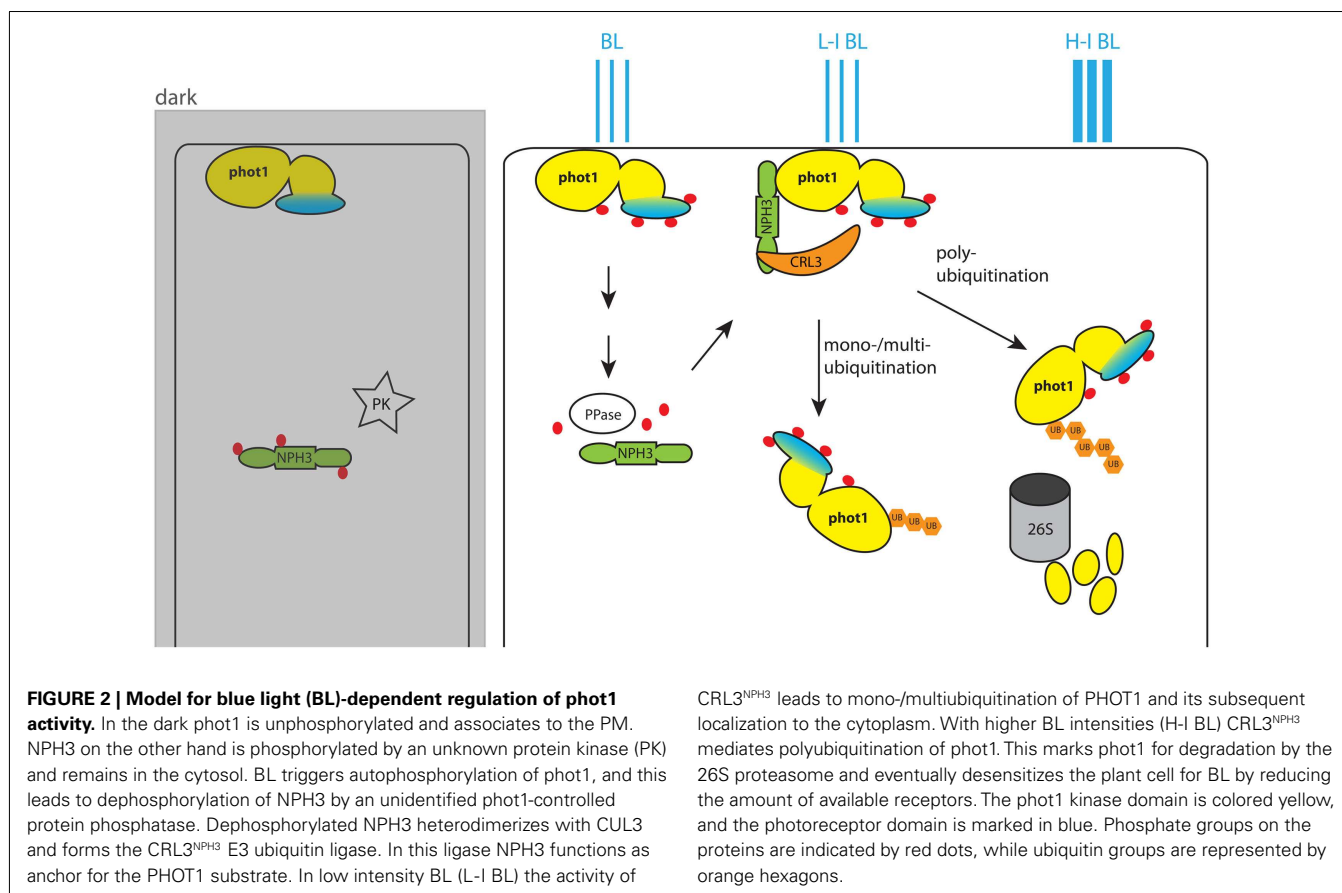
## DYNAMIC LOCALIZATION OF AGC KINASES

Apart from being activated, protein kinases need to be brought into proximity of their substrates. Vice versa, separation of the kinase from its substrates by subcellular re-localization is an efficient mechanism to regulate kinase activity. AGC protein kinases phosphorylate nuclear, cytosolic, or membrane localized target proteins, and their proper localization relies on internal localization signals or on binding to scaffold proteins, or specific membrane components.

An example of re-localization in animal kinases is the current model for the previously discussed Akt/PKB activation by PDK1. PDK1 can change from a cytosolic homodimer into a monomer that binds Akt/PKB. Association of this heterodimer through the PH domains of both proteins to the PM triggers PDK1 to phosphorylate Akt/PKB, and the subsequent release of active Akt/PKB. The monomeric PDK1 then dissociates from the membrane to form a “primed” homodimer, or to retrieve the next Akt/PKB target protein (Masters et al., 2010).

In plants the activity of the phototropins seems also to be regulated by re-localization. In dark grown seedlings phot1 and 2 localize to the PM. A pulse of BL induces partial endocytosis of phot1 (Sakamoto and Briggs, 2002) and association of phot2 with the Golgi apparatus (Kong et al., 2006). Recent work suggests that internalization of phot1 is regulated by the ubiquitination status of the kinase (Roberts et al., 2011), the mechanism of which will be further discussed below (see Section “Plant AGC Kinase Activity Dynamics in Development and Growth”, Figure 2).

Similarly, investigation of a complementing PID::VENUS fusion revealed that PID is mainly present at the PM, close to its phosphorylation targets the PIN proteins (Michniewicz et al.,



2007). That PM localization is important for PID action was indicated by the reduced effect of PID overexpression in a sterol biosynthesis mutant background (Dhonsukshe et al., 2010). In this mutant background PID:VENUS was found to localize predominantly to the cytosol, indicating that the membrane composition is important for PID PM localization, and that PID PM localization is important for PID function. PID can also be found in punctuate structures in the cytoplasm (Michniewicz et al., 2007; Dhonsukshe et al., 2010), which is intriguing, since the only demonstrated function of PID is phosphorylation of PIN family members at the PM. It is tempting to speculate that these cytoplasmic structures are either part of a desensitization mechanism and involved in PID turn-over, or that PID has other phosphorylation targets in the cytosol.

In maize the PID co-ortholog BARREN INFLORESCENCE2 (BIF2) has been identified. Like PID, a YFP:BIF2 fusion localizes to the PM and phosphorylates the HL of ZmPIN1a *in vitro* (Skirpan et al., 2008, 2009). This is necessary for proper localization of ZmPIN1a in developing inflorescence meristems (Skirpan et al., 2009), and suggests that phosphorylation-directed control of PIN protein targeting by AGC kinases is a conserved regulatory mechanism, at least in seed plants. Localization studies in onion epidermis and *Nicotiana benthamiana* cells have revealed that YFP:BIF2 is also present in the nucleus, where it seems to interact with and to phosphorylate the basic helix-loop-helix transcription factor BARREN STALK1 (BA1). Nuclear localization of BIF2 might be mediated through its interaction with BA1 or through a

yet unidentified nuclear localization signal (Skirpan et al., 2008). Nuclear localization has not been reported for PID, however, two other AGC3 kinases WAG1 and WAG2 seem to act similar to BIF2, in that they phosphorylate PINs at the PM (Dhonsukshe et al., 2010) and localize to the nucleus (Galvan-Ampudia and Offringa, 2007) where they possibly regulate the activity of nuclear proteins.

The actual protein domains involved in PM association of phot1/2 and PID are not known. A transmembrane domain is clearly absent in these proteins, implying that PM association is achieved by interaction with other PM-bound proteins or through binding to lipid components of the PM. Binding of PID to several phosphoinositides and phosphatidic acid (PA) in a lipid-overlay assay suggested that the latter is true. A fusion of GFP with the insertion domain of PID localizes to the PM in yeast cells, demonstrating that this domain is sufficient for PM localization. This is further substantiated by the redirection of AGC1–7 from the cytoplasm to the PM in tobacco cells after exchange of its insertion domain for the PID insertion domain (Zegzouti et al., 2006b). Despite these indications, mechanisms that regulate subcellular localization of PID and the other AGC3 kinases remain elusive.

*Arabidopsis* PDK1 was also found to bind to phosphoinositides and PA in lipid-overlay assays through a PH domain present at its C-terminus (Deak et al., 1999). Importantly, only binding to PA stimulates PDK1 activity in a kinase assay (Anthony et al., 2004; Otterhag et al., 2006), suggesting that PDK1 activity might be restricted to specific (PA-containing) locations at the PM.

## REGULATION BY INTERACTING PROTEINS AND SECOND MESSENGERS

Apart from regulation of AGC protein kinase activity by other kinases and re-localization of the kinases and their substrates, only a few other regulatory factors have been identified.

Mammalian PDK1 is negatively regulated by the 14-3-3 protein  $\theta$ . A similar role has been indicated for 14-3-3 proteins in *Arabidopsis* by assaying the phosphorylation of S6K2 by PDK1 in the presence of 12 14-3-3 isoforms. In this assay autophosphorylation of PDK1 and subsequent phosphorylation of S6K2 was enhanced by 9 14-3-3s ( $\mu$ ,  $\phi$ ,  $\kappa$ ,  $\omega$ ,  $\epsilon$ ,  $\psi$ ,  $\nu$ ,  $\chi$ ,  $\nu$ ) and completely inhibited by another ( $\sigma$ ; Otterhag et al., 2006). Since in all cases phosphorylation levels of PDK1 were changed by the 14-3-3 proteins, it was suggested that the 14-3-3 proteins facilitate dimerization of PDK1 and that this enhances the activating trans-autophosphorylation of PDK1. Recent reports also indicate a role for 14-3-3 proteins in the regulation of phot1 and 2. An interaction between phot1 and the 14-3-3 proteins  $\lambda$ ,  $\kappa$ ,  $\phi$ ,  $\nu$  was demonstrated in yeast-two-hybrid assays and by immunoprecipitation from plant extracts (Inoue et al., 2008; Sullivan et al., 2009). Interestingly, the interaction of phot1 with 14-3-3  $\lambda$  was found to be light dependent in both assays, but a role for this interaction was not investigated (Sullivan et al., 2009). Phot2 on the other hand was found to interact with 14-3-3  $\lambda$  in a yeast-two-hybrid assay. Further investigation of 14-3-3  $\lambda$  binding to fragments of phot2 demonstrated that the kinase domain is sufficient for this interaction. Furthermore, replacement of a serine in the 14-3-3 binding site (RSK[S→A]QP) located in the T-loop of PHOT2 was sufficient to abolish this interaction in yeast. *In planta* loss of 14-3-3  $\lambda$  reduces phot2 functionality only in BL induced stomatal opening, while other phot2 regulated processes are not affected (Tseng et al., 2012). In conclusion, 14-3-3 proteins seem to interact with AGC kinases and to influence their activity. Whether this is by enhancing protein interactions for trans-autophosphorylation or substrate phosphorylation, or by stabilizing the active conformation of AGC kinases has yet to be determined.

Together with interacting proteins, second messengers play an essential role in the activation of several AGC kinases. Earlier we described the dependence of PDK1 activation on interaction with 3-phosphoinositides at the PM (see Sections “PDK1 as Regulator of AGC Kinase Activity” and “Dynamic Localization of AGC Kinases”). In a similar manner binding of cyclic guanosine monophosphate (cGMP) is necessary for the activation of isoforms of the cyclic nucleotide-dependent kinase PKG. Here, binding of cGMP initiates conformational changes that relieve the kinase from an autoinhibitory domain. Isoforms of PKA, protein kinase X (PRKX), and PRKY are controlled by cyclic adenosine monophosphate (cAMP), but in the off state these kinases exist as hetero-tetramers of two catalytic (C-) and two regulatory (R-) subunits. When G-protein coupled receptors activate adenylate cyclase, the produced cAMP binds to the R subunits, causing dissociation of the C subunits, and activation by phosphorylation of their activation loop (reviewed in Pearce et al., 2010). Even though cAMP and cGMP based signaling has been shown to be important in plant development and stress responses, the existence of kinases that respond to concentration changes of these second messengers remains elusive (Isner et al., 2012).

Another well-known mechanism is calcium-dependent regulation of protein kinase activity. A plant- and protozoan-specific class of calcium-regulated kinases are the Calmodulin-like Domain Protein Kinases (CDPKs) that have their own calmodulin (CaM) domain with four calcium binding pockets (EF-hands) to sense the calcium concentration. Binding of  $\text{Ca}^{2+}$  to the CaM domain relieves the repression by the autoinhibitory junction domain and thus leads to kinase activation (Hrabak et al., 2003). Other kinases only have a CaM binding domain and are therefore named CaM dependent protein kinases (CaMKs). CaMKs are activated by CaM binding, and are found in animals, but have only occasionally been identified in plants (Carafoli et al., 2001). A plant-specific group of CaMKs that has been initially identified in lily is the calcium and CaM-dependent protein kinase (CCaMK; Hrabak et al., 2003). Besides a CaM binding domain, CCaMKs have a visinin-like C-terminal domain with three EF-hands. A well-studied example of the CCaMKs is DMI3, which plays a central role in the symbiotic interaction of legumes with rhizobia and arbuscular mycorrhizal fungi. In both cases DMI3 translates the micro-organism-induced calcium spiking into respectively root nodule and arbuscular mycorrhiza formation. The EF-hands in the C-terminal domain seem to regulate kinase autophosphorylation at basal calcium levels, which in turn enhances binding of the CaM at calcium peaks, relieving auto-inhibition, and leading to activation of the kinase and substrate phosphorylation (Sathyanarayanan et al., 2000; Gleason et al., 2006; Swainsbury et al., 2012). CaM-dependent kinases are only found in certain plant species, and neither CaMK, nor CCaMK can be found in *Arabidopsis* (Hrabak et al., 2003).

Regulation by  $\text{Ca}^{2+}$  has been reported for the conventional animal PKC isoforms. Calcium binding to the C2 domain enhances PM association and subsequent activation of PKCs (Newton, 2001). To our knowledge regulation of AGC kinases through direct binding of CaMs has not been reported in animal systems. In contrast, the *Arabidopsis* PID kinase activity is regulated by the small calcium binding protein PID-BINDING PROTEIN 1 (PBP1) and the CaM-like protein TOUCH3 (TCH3). Binding of these proteins to PID is enhanced by calcium. However, while PBP1 interacts weakly with PID in the absence of  $\text{Ca}^{2+}$ , binding of TCH3 is strictly  $\text{Ca}^{2+}$ -dependent. Importantly, autophosphorylation of PID is enhanced by addition of PBP1 in the kinase assay, while it is repressed by TCH3 (Benjamins et al., 2003). There is some analogy in the regulation of PID activity by two types of calcium binding proteins and the calcium-dependent regulation of the CCaMK DMI3 (Swainsbury et al., 2012), which leads to the interesting hypothesis that PBP1 and TCH3 might allow modulation of PID kinase activity in response to oscillating cytosolic calcium levels.

Previously, it was reported that PID is directly inhibited *in vitro* by 15 mM  $\text{Ca}^{2+}$ . Because PID does not have an obvious  $\text{Ca}^{2+}$  binding pocket, and co-incubation with 15 mM  $\text{Ca}^{2+}$  and  $\text{Mg}^{2+}$  restores PID activity, this inhibition is likely to be a result from competitive replacement of the  $\text{Mg}^{2+}$  ion in the catalytic core by  $\text{Ca}^{2+}$  (Zegzouti et al., 2006a). Hence, it might be that AGC kinase activity is regulated by the availability of mandatory cofactors such as  $\text{Mg}^{2+}$  and ATP. The chance, however, that  $\text{Ca}^{2+}$  will



be inhibiting PID *in vivo* is low, as reported cytosolic  $Mg^{2+}$  concentrations are in the millimolar range, whereas those for  $Ca^{2+}$  range from sub-micromolar to micromolar (Hepler, 2005).

Finally, evidence for direct binding of the natural kinase inhibitor quercetin to PID was recently provided (Henrichs et al., 2012). Other than the kinase inhibitors staurosporine and chelerythrine, the flavonol quercetin is able to efficiently inhibit substrate phosphorylation by PID. This coincides with the reported function of quercetin as a natural polar auxin transport inhibitor (Jacobs and Rubery, 1988). Both PID expression and flavonoid biosynthesis are upregulated in response to auxin, PID expression within 2 h, and flavonoid accumulation only 8–12 h after auxin treatment (Benjamins et al., 2001; Lewis et al., 2011). The flavonol quercetin could thus provide feedback control on auxin enhanced PID kinase activity.

## PLANT AGC KINASE ACTIVITY DYNAMICS IN DEVELOPMENT AND GROWTH

In the previous sections we have described important structural features of plant specific AGC protein kinases as well as known regulatory events that directly act on these protein kinases to modulate their activity. While this might help in understanding AGC protein kinase functionality, many regulatory pathways that have not been directly connected to the kinases had to be left out. Here we will use the modulation of auxin responses and transport by on the one hand the most ancestral AGCVIII kinases phot1 and phot2, and on the other hand the land plant specific AGCVIII kinases PID, WAG1, and WAG2 to provide examples of how all aspects of AGC kinases are integrated in their role as regulators of plant development.

### REGULATING PHOTOTROPIC GROWTH

Phot1 and phot2 regulate various processes in response to BL. Both kinases have been shown to regulate phototropism, stomatal opening, chloroplast accumulation movement, and cotyledon/leaf flattening. Additionally, a role of both proteins in regulating leaf movement has been implicated. Next to these shared roles in which both phototropins only distinguish themselves by acting at respectively lower and higher light fluence rates, they also have unique regulatory roles. Phot1 has been shown to induce hypocotyl growth inhibition and to be involved in the regulation of mRNA stability, whereas chloroplast relocation to avoid intense light solely depends on phot2 (Christie, 2007).

For all these processes various factors were identified that act downstream of phot1/phot2-mediated phosphorylation. Yet apart from BL of different intensities, only a few mechanisms were indicated in fine tuning the activity of one or both phototropins. As mentioned earlier, phototropins locate to the PM in dark grown seedlings and are relocated to cytoplasmic- or Golgi-like structures after BL treatment (Sakamoto and Briggs, 2002; Kong et al., 2006). For phot1, internalization has been suggested to be modulated by ubiquitination through the CULLIN-RING E3 ubiquitin ligase CRL3<sup>NPH3</sup> to which it binds via the substrate adaptor NON-PHOTOTROPIC HYPOCOTYL 3 (NPH3) (Roberts et al., 2011). Interestingly, in dark grown seedlings NPH3 is found to be phosphorylated. Yet a BL pulse leads to its dephosphorylation in a phot1-dependent manner (Pedmale and Liscum, 2007).

Neither the kinase that phosphorylates NPH3 in the dark nor the protein phosphatase that dephosphorylates it in response to phot1 activation have been identified to date (Figure 2). Possibly, dephosphorylation enhances the affinity of NPH3 for phot1, and at the same time stabilizes the assembly of CRL3<sup>NPH3</sup>. Roberts and coworkers propose a model in which the degree of phot1 ubiquitination is depending on BL intensity (Figure 2): under low intensity BL phot1 would be mono-/multi-ubiquitinated and potentially internalized into endosomes via a clathrin-dependent mechanism (Kaiserli et al., 2009; Roberts et al., 2011). Higher BL intensities would lead to polyubiquitination of phot1 and its subsequent recruitment for degradation by the 26S proteasome, thereby desensitizing cells for BL (Roberts et al., 2011).

A phot2:GFP fusion was shown to interact with the PM through its kinase domain, and to co-localize with a Golgi marker in a punctate pattern in the cytosol few minutes after its activation by BL treatment (Kong et al., 2006). A strikingly similar punctate pattern was observed for the osmotic stress-related protein kinase SnRK2.4 a few minutes after application of salt stress (McLoughlin et al., 2012). It is tempting to speculate that these are similar subcellular structures involved in rapid feed back regulation of kinase activity. *Chlamydomonas* phot2 can complement the *Arabidopsis* mutant (Onodera et al., 2005), and its activation was proposed to lead to a conformational change of the insertion domain (Pfeifer et al., 2010). If we assume that, analogous to PID, the insertion domain is responsible for PM association, this conformational change might thus be responsible for dissociation or trafficking from the PM.

Next to NPH3, various other proteins have been shown to interact with the phototropins (for review see Inoue et al., 2010). Of these only ATP BINDING CASSETTE B19 (ABCB19) and recently PHYTOCHROME KINASE SUBSTRATE 4 have been identified as substrate of phot1 (Christie et al., 2011; Demarsy et al., 2012).

In protein extracts from 3-day-old etiolated seedlings phosphorylated PKS4 (PKS4L) could be detected after 30 s of BL irradiation. A maximum of PKS4L was detectable after 10 min of irradiation, after which PKS4L levels decreased and finally were no longer detectable after 4 h of irradiation. PKS4L was detectable in *phot2* mutants but absent in mutants lacking phot1 or expressing a kinase dead versions of it. In addition, a truncated version of PHOT1 was found to phosphorylate several fragments of PKS4 *in vitro*. This demonstrates that PKS4 is a substrate of phot1. Accordingly, accumulation of PKS4L could be enhanced either by irradiation with BL of higher intensities, or by treatment with a phosphatase inhibitor. Measurement of phot1-mediated phototropic bending in response to BL indicated that unphosphorylated PKS4 positively regulates phototropism, while this response is inhibited by PKS4L (Demarsy et al., 2012). Although PKS4 is likely to be part of a negative feedback loop on phot1 activity, a direct effect of this protein on phot1 activity remains to be demonstrated.

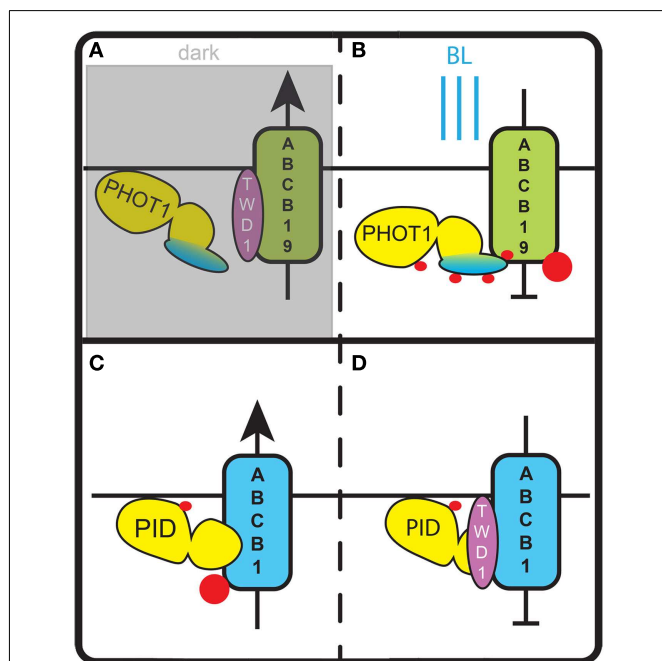
ABCB19 is an auxin efflux carrier that functions in the long distance transport of auxin from the shoot apical meristem to the root, and the immunophilin TWISTED DWARF1 (TWD1) has been suggested to act as a positive modulator of its activity (Bouchard et al., 2006). Binding of ABCB19 and its phosphorylation by phot1 was demonstrated *in vitro* and in yeast cells. In HeLa cells co-transfected with ABCB19 and phot1 auxin efflux is abolished after BL treatment, indicating that phosphorylation

of ABCB19 by phot1 inhibits the auxin transporter. Further immuno-precipitation assays showed that the interaction between TWD1 and ABCB19 is lost after BL treatment of wildtype seedlings, but can be detected in equally treated *phot1* seedlings. This indicates that phot1-dependent phosphorylation of ABCB19 disrupts the positive modulation of ABCB19 by TWD1 at the light side of the hypocotyl (Christie et al., 2011; **Figures 3A,B**). The subsequent PIN mediated redirection of auxin flow toward the shaded hypocotyl side (Ding et al., 2011) would stimulate cell expansion on that side and thus result in phototropic bending in response to a directional BL stimulus.

### REGULATING POLAR AUXIN TRANSPORT

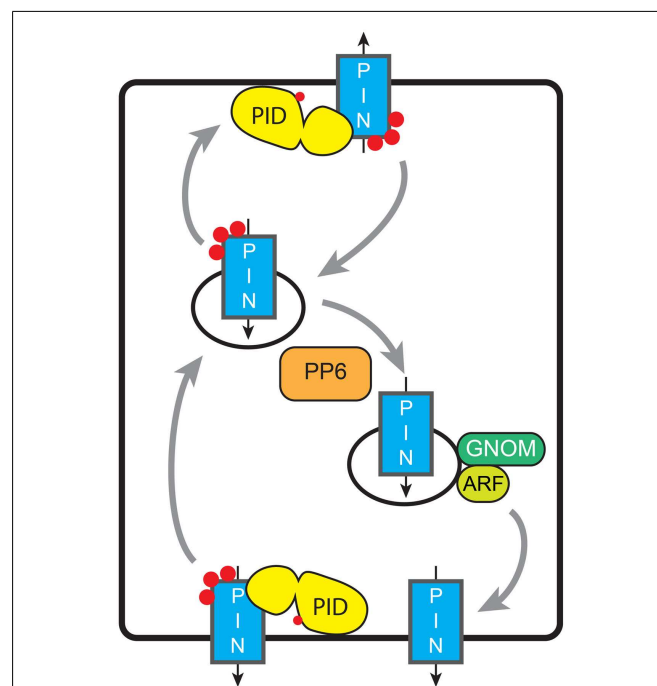
The studies on the phototropins have revealed how their activity is regulated by light. A few components in the respective downstream signaling pathways, such as for example NPH3 (Roberts et al., 2011), PKS4 (Lariguet et al., 2006), and ABCB19 (Christie et al., 2011), have been uncovered. However, the signaling processes downstream of phototropins are only now beginning to be unraveled. In contrast, studies on the redundantly acting *Arabidopsis* AGC3 kinases PID, WAG1, and WAG2 have revealed the PIN auxin efflux carriers as one of their main targets

(Michniewicz et al., 2007; Dhonukshe et al., 2010; Huang et al., 2010), while the exact functions of identified proteins that bind to PID or its maize ortholog BIF2 remain unclear (Benjamins et al., 2003; Skirpan et al., 2008). The three *Arabidopsis* AGC3 kinases were shown to phosphorylate serine residues in three conserved TPRxS[N/S] motifs in the HL of PIN1-type auxin efflux carriers, leading to apical (shootward) PIN localization, whereas loss-of-phosphorylation was shown to induce a PIN polarity shift toward the basal (rootward) cell membrane (Friml et al., 2004; Dhonukshe et al., 2010; Huang et al., 2010). The three AGC3 kinases act antagonistically in determining PIN polarity with a novel PP6-type heterotrimeric protein phosphatase, consisting of a PP2AA regulatory subunit, the PP6 catalytic subunit FyPP1 or FyPP3, and the SAPS DOMAIN-LIKE protein SAL that binds to the PIN-HL in a phosphorylation-dependent manner (Michniewicz et al., 2007; Dai et al., 2012; **Figure 4**). Interestingly, several other serine and threonine residues in the PIN-HL have been identified as *in vivo* phosphorylation targets (Nuhse et al., 2004; Benschop et al., 2007; Michniewicz et al., 2007; Chen et al., 2010). For at least two of these residues in the PIN1-HL it was shown that their phosphorylation



**FIGURE 3 | Regulation of ABCB auxin transporters by AGC kinases.**

**(A)** In darkness phot1 is inactive and does not phosphorylate ABCB19. In this state ABCB19 can interact with its positive modulator TWD1 and export auxin from the cell. **(B)** Blue light (BL) activates phot1 and induces phosphorylation of ABCB19. This inhibits ABCB19 – TWD1 interaction and blocks auxin efflux via ABCB19. **(C)** In the absence of TWD1, phosphorylation of a serine in the linker domain of ABCB1 by PID stimulates ABCB1 activity and leads to an increase of auxin export. **(D)** In the presence of TWD1, PID is no longer able to phosphorylate the linker domain of ABCB1. This lack of phosphorylation and potentially the phosphorylation of other sites of ABCB1 block ABCB1 activity and the respective auxin efflux. Red dots indicate phosphorylation events.



**FIGURE 4 | Model for phosphorylation-dependent re-localization of PINs.**

De novo synthesized PINs are distributed apolarly to the PM. Constitutive endocytosis relocates PINs depending on their phosphorylation status (red dots). This relocation might be baso-apical as in the figure (embryo, root, shoot, and inflorescence meristem), apolar-lateral (phototropism), between lobe and indentation (pavement cells), or between the inner or outer cell side (guard cells). Tuning of the activities of AGC kinases and PP6-type phosphatase determines the PIN phosphorylation status and thus the preferential targeting pathway, and leads to the respective redistribution of the PINs. Basal targeting of unphosphorylated PINs is GNOM-dependent; ARF-GEFs involved in other targeting pathways remain unknown. Grey arrows indicate trafficking of PIN-loaded vesicles, while black arrows indicate the direction of PIN-mediated auxin transport.

status determines the asymmetric distribution of PIN1 (Zhang et al., 2010). It is likely that these other residues are phosphorylated by other kinases that remain to be identified. Potential candidates are the four D6 and D6-like protein kinases of the AGC1 group (AGC1-1/D6PK, AGC1-2/D6PKL1, PK5/D6PKL2, PK7/D6PKL3). These kinases were shown to affect auxin transport, possibly by phosphorylating the PIN-HL, but their activity does not lead to changes in PIN polarity (Zourelidou et al., 2009; Dhonukshe et al., 2010). It will be interesting to see whether these kinases recognize the same or different residues in the PIN-HL, or that they act on auxin transport via a different mechanism.

An important question that remains to be answered is: what in the cell recognizes the phosphorylation status of PIN proteins to direct their proper localization at the PM? Studies with PIN-fluorescent protein fusions indicate that *de novo* synthesized PIN proteins are initially placed on the PM in an apolar manner, where most likely their phosphorylation status is determined by the competing activities of PM-associated AGC3 kinases and PP6-type phosphatases (Dhonukshe et al., 2010). Following clathrin- and sterol-dependent endocytosis (Dhonukshe et al., 2007; Men et al., 2008), the PIN-HL phosphorylation status is somehow sensed, resulting in recycling to the PM at the correct side of the cell. This dynamic trafficking of PIN-loaded vesicles has been shown to involve the actin cytoskeleton (Geldner et al., 2001), and the action of ADP Ribosylation Factors (ARFs; Xu and Scheres, 2005), their downstream Rho GTPases (ROPs; Xu et al., 2010; Nagawa et al., 2012) and their upstream ARF GTP exchange factors (ARF-GEFs; Geldner et al., 2001, 2003; Sieburth et al., 2006; Kleine-Vehn et al., 2008). Long term inhibition of the ARF-GEF GNOM by low concentrations of the fungal toxin brefeldin A (BFA) was also found to lead to a basal-to-apical transcytosis of PIN proteins, indicating that GNOM is involved in recycling of PIN proteins to the basal PM, whereas other ARF-GEFs induce trafficking to the apical membrane (Geldner et al., 2003; Kleine-Vehn et al., 2008). The phosphorylation status of the PIN-HL was shown not to inhibit PIN endocytosis (Dhonukshe et al., 2010), but rather to determine the recruitment of PIN proteins into the GNOM-independent trafficking pathway (Kleine-Vehn et al., 2009). The phosphorylation signal thus directs super-polar PIN targeting during recycling, whereas maintenance of PIN polarity by recruitment of PINs into non-mobile PM clusters and by spatially defined endocytosis (Kleine-Vehn et al., 2011) seems to be independent of the PIN phosphorylation status. AGC3 kinase-mediated shifts in PIN polarity have been shown to be important for tropic growth responses (Sukumar et al., 2009; Dhonukshe et al., 2010; Ding et al., 2011), and for patterning during embryogenesis and organ formation at the shoot and inflorescence meristems (Kleine-Vehn et al., 2009; Huang et al., 2010; Li et al., 2011). However, not for all processes changes in AGC3 kinase activity lead to the predicted apico-basal shift in PIN localization. For example, the phototropic response of the hypocotyl of etiolated *Arabidopsis* seedlings was shown to be mediated by BL dependent relocation of the PIN3 auxin efflux carrier (based on a complementing *PIN3::PIN3:GFP* fusion) from a symmetric distribution to an asymmetric localization at the inner lateral side of hypocotyl endodermis cells. Genetic evidence was provided that at least the PID kinase is involved in this response. The observed phot1-dependent down regulation of *PID*

expression was proposed to lead to PIN3 loss-of-phosphorylation and thus to GNOM dependent trafficking of PIN3 proteins to the inner lateral side (Ding et al., 2011). How the phototropin signal is translocated to the nucleus to affect PID transcription, and whether the phot-dependent rapid increase in cytosolic  $\text{Ca}^{2+}$  concentration following BL stimulation (Baum et al., 1999; Babourina et al., 2002; Harada et al., 2003) has a role in these responses is unclear. The possibility that the phot1 kinase directly phosphorylates PIN proteins was excluded by *in vitro* phosphorylation reactions (Ding et al., 2011), indicating that phototropins and AGC3 kinases have different substrate specificities.

The PID/PP6-dependent phosphorylation status of PIN1 was also shown to modulate the interdigitated pattern of epidermis cells in *Arabidopsis* leaves. PID overexpression or PP6 loss-of-function induced a shift of PIN1:GFP localization to the indentations, leading to a reduction in the degree of pavement cell indentation, whereas phosphatase overexpression reversed this to wildtype PIN1:GFP localization in the lobe (Li et al., 2011). PID overexpression or PP6 loss-of-function were also shown to induce re-localization of PIN1:GFP from the inner to the outer side of stomatal guard cells (Li et al., 2011). In conclusion, depending on the cell- and tissue type there are variations on the initial paradigm that AGC3 kinase-mediated phosphorylation of PIN proteins directs their baso-apical polarity. The central message is that *de novo* synthesized PIN proteins come into contact with these kinases upon arrival at the PM, and that phosphorylation leads to their sorting in the GNOM-independent pathway, whether this is apical in meristem epidermis cells, lateral-outer in hypocotyl endodermis cells, or outer in stomatal guard cells. The PM association of PID close to its phosphorylation targets is central here, and it will be interesting to map the site that allows association with the PM. In addition it will be important to establish how the calcium binding PID interacting proteins affect PID activity in response to changes in the cytosolic calcium concentration.

A recent study provides evidence that PID does not only phosphorylate PIN auxin efflux carriers, but also acts on an auxin transporter of the ABCB family (Figures 3C,D). ABCB auxin transporters are predominantly apolarly localized in plant cells and have been proposed to reduce net-influx of auxin from the apoplast (Mravec et al., 2008). Evidence was obtained that PID modulates the auxin transport activity of ABCB1 by phosphorylating a serine residue in the linker domain of this transporter (Henrichs et al., 2012). It is tempting to speculate that this occurs equivalent to the modulation of ABCB19 activity by phot1, and that accordingly ABCB1 phosphorylation by PID disrupts the interaction with TWD1 (Figure 3). However, measurements of auxin efflux from *Nicotiana benthamiana* protoplasts co-transfected with PID and ABCB1 showed increased efflux, whereas co-transfection of ABCB1 and TWD1 led to a reduction in efflux. Co-transfection of PID, ABCB1, and TWD1 in the same system completely abolished auxin efflux. In yeast cells, the contradictory finding that ABCB1 activity is reduced by PID co-expression might be explained by a yeast immunophilin (ScFKBP12) that acts redundantly to TWD1 (Henrichs et al., 2012). Taken together, this indicates that PID-dependent phosphorylation of ABCB1 has an activating or inhibiting effect depending on the absence or presence of TWD1 (Figures 3C,D). In this way, TWD1 might control the rate of

polar auxin transport, by serving as a switch between asymmetric polar auxin efflux by PIN family transporters and symmetric auxin efflux by ABCB1 (Mravec et al., 2008). Interestingly, the effect of PID-mediated phosphorylation differs per type of transporter: for PIN proteins it directs subcellular localization, whereas for ABCB1 it affects auxin transport activity (Figures 3 and 4).

## CONCLUSION

Plants share the basic AGC kinase subfamilies (PDK1, S6Ks, and NDRs) with other eukaryotes, but lack the typical PKA and PKC subfamilies that are involved in regulating cell growth, -division, and -polarity in animals and yeast. Instead, plants seem to have adopted the AGCVIII kinase subfamily to perform similar functions. The first AGCVIII kinase appeared in unicellular algae as BL-sensing photoreceptor (phot2), whose signaling protects the cells against high fluency rate BL. During the evolution of multicellular plant life and the move from water to land, the AGCVIII kinases diversified by losing the photo-sensing domain, and were recruited for other functions such as control of auxin transport (PID, WAG1, WAG2, D6PK, and D6PK-L) and pathogen responses (OXI1) (Galvan-Ampudia and Offringa, 2007). Even though the vital role of these kinases for proper plant development is now well-established, not much is known about their regulation and only a few substrates have been identified to date.

The strong conservation of the catalytic kinase core among AGC kinases implies that functional and structural knowledge obtained by detailed studies on mammalian AGC kinases also applies to plant AGC kinases. For example, two basic regulatory processes identified in animal kinases were later also demonstrated for plant AGC kinases: (i) the necessity of T-loop phosphorylation for full activation, and (ii) the potential involvement of PDK1 in this activation, including its interaction via the C-terminal PIF domain. It remains to be determined, however, how plant AGC

kinases are activated. With PDK1 a general upstream activator of AGC kinases in plants might be present, but at least for the phototropin kinase domains and several other AGC kinases (PID, WAG1, WAG2, OXI1) auto-activation has been reported. For PID, activation was stronger in the presence of PDK1 (Zegzouti et al., 2006a,b). Other upstream kinases that phosphorylate the T-loop of AGC kinases have not yet been identified.

Next to activation of the T-loop we have highlighted the potential regulatory domains in the insertion domain of the activation segment and in the N- and C-tail that extend from the kinase core of plant AGC kinases. As in animal AGC kinases, the relatively large N- and C-tails of plant AGC kinases suggest that protein-protein interaction domains may be found here. The insertion domain that is typical for the plant specific AGCVIII kinases was found to be essential for proper subcellular localization of *Ara-bidopsis* PID and tomato Adi3. Although this may only be one of the functional aspects of the insertion domain, it is likely that also other plant AGC kinases have a signaling sequence in their insertion domain that guides them to their proper location. One might assume that the conformation of this signaling sequence changes depending on T-loop phosphorylation status and the interaction with regulatory proteins. This would give the cell a straightforward mechanism to control the localization of activated AGC kinases. Several AGCVIII kinases have been shown to be present at multiple locations within the cell (e.g., phototropins, WAG1, WAG2, ZmBIF2). It will be interesting to see which intrinsic signals instruct their subcellular localization, and whether this is modulated in response to distinct stimuli. Further dissection of the AGC kinase subdomains guided by the structural knowledge presented in this review will help to understand the dynamics of intracellular localization of plant AGC kinases in relation to the phosphorylation state of their targets and the processes that they regulate.

**Table 2 | AGC kinases characterized in other plant species.**

Kinase name	Species	Function	<i>Arabidopsis</i> homolog	Literature
Adi3	<i>Solanum lycopersicum</i>	Cell death suppression	AGC1-3	Devarenne et al. (2006)
BARREN	<i>Zea mays</i> , <i>Oryza sativa</i> , <i>Pisum sativum</i>	Auxin transport	PID, WAG1, WAG2	Bai et al. (2005), McSteen et al. (2007), Morita and Kyo-zuka (2007)
INFLORESCENCE 2 (BIF2), OsPID, PsPK2				
IRE	<i>Medicago truncatula</i>	Nodule formation	IRE	Pislaru and Dickstein (2007)
PDK1	<i>Oryza sativa</i> , <i>Physcomitrella patens</i>	AGC kinase activation	PDK1	Matsui et al. (2010), Dittrich and Devarenne (2012)
Phototropins	<i>Avena sativa</i> , <i>Chlamydomonas reinhardtii</i> , <i>Oryza sativa</i> , <i>Pisum sativum</i> , <i>Vicia faba</i> , etc.	Blue light perception	PHOT1/2	Phylogenetic overview in Lariguet and Dunand (2005)
PKV	<i>Solanum lycopersicum</i>	Disease resistance	AGC1-7	Hammond and Zhao (2000)
PKL01	<i>Lotus japonicus</i>	n. d.	NDR	Kameshita et al. (2010)
OXI1	<i>Oryza sativa</i>	Disease resistance	OXI1/AGC2-1	Matsui et al. (2010)
CsPK3, PsPK3	<i>Cucumis sativus</i> , <i>Pisum sativum</i>	Auxin transport	WAG1, WAG2	Santner and Watson (2006)

Homologs of various *Arabidopsis* AGC kinases have been described in several plant species.

The table respectively indicates the name of the kinase, the plant species in which it has been identified, the biological process in which the kinase functions, the corresponding *Arabidopsis* homologs, and literature references. Homologs of phototropins have been identified in a plethora of different plant species. For initial reading we refer here to a review. The regulatory function of PKL01 in *Lotus japonicus* has not been determined yet. A recent report indicates that it might be able to phosphorylate tyrosine as well as serine and threonine residues.



The identification of novel AGC kinases can be surprising as a recent report on *Lotus japonicus* PKL01, an NDR kinase (AGCVII) homolog, describes this AGC kinases as being able to phosphorylate tyrosine residues next to its conventional serine/threonine kinase activity (Katayama et al., 2012). To our knowledge the function of AGCVII kinases in *Arabidopsis* has not been determined yet. It will be interesting to see if this expanded function is present in *Arabidopsis*, and which structural changes might enable the dual-specificity of the NDR kinases.

Furthermore, homologs of *Arabidopsis* AGC kinases have been characterized in other plant species (see Table 2). Next to demonstrating similar roles of some AGC kinases in different plant species, these studies also revealed interesting differential aspects of AGC kinase functionality. For example, in contrast to the reported conserved functions of several AGCVIII kinases, INCOMPLETE ROOT HAIR ELONGATION (IRE) a kinase of the “AGC other” group seems to have acquired a new function in *Medicago truncatula*. In *Arabidopsis* this kinase has been shown to regulate root hair elongation (Oyama et al., 2002), while in *Medicago* a role in the

formation of nodules has been described (Pislariu and Dickstein, 2007). Whether this is due to the lack of distinct downstream targets in *Arabidopsis*, or due to a changed role of IRE in *Medicago* remains unknown, and mis-expression experiments with the two proteins as well as identification of substrates in the respective plants will be needed.

In conclusion, plant AGC kinases are an interesting group of kinases that perform functions conserved in all eukaryotes, but also plant-specific functions. Especially the plant-specific AGCVIII kinases have several interesting features, such as their dynamic subcellular localization, and we are confident that further dissection of their structure and function will lead us to new paradigms in signaling and cell polarity regulation.

## ACKNOWLEDGMENTS

Eike H. Rademacher was supported by a TOP grant from the Research Council for Chemical Sciences (700.58.301) to Remko Offringa, with financial aid from the Netherlands Organization of Scientific Research (NWO).

## REFERENCES

- Anthony, R. G., Henriques, R., Helfer, A., Meszaros, T., Rios, G., Testerink, C., et al. (2004). A protein kinase target of a PDK1 signalling pathway is involved in root hair growth in *Arabidopsis*. *EMBO J.* 23, 572–581.
- Babourina, O., Newman, I., and Shabala, S. (2002). Blue light-induced kinetics of H<sup>+</sup> and Ca<sup>2+</sup> fluxes in etiolated wild-type and phototropin-mutant *Arabidopsis* seedlings. *Proc. Natl. Acad. Sci. U.S.A.* 99, 2433–2438.
- Bai, F., Watson, J. C., Walling, J., Weeden, N., Santner, A. A., and Demason, D. A. (2005). Molecular characterization and expression of *PSPK2*, a PINOID-like gene from pea (*Pisum sativum*). *Plant Sci.* 168, 1281–1291.
- Balendran, A., Biondi, R. M., Cheung, P. C., Casamayor, A., Deak, M., and Alessi, D. R. (2000). A 3-phosphoinositide-dependent protein kinase-1 (PDK1) docking site is required for the phosphorylation of protein kinase C $\zeta$  (PKC $\zeta$ ) and PKC-related kinase 2 by PDK1. *J. Biol. Chem.* 275, 20806–20813.
- Bastidas, A. C., Deal, M. S., Steichen, J. M., Keshwani, M. M., Guo, Y., and Taylor, S. S. (2012). Role of N-terminal myristylation in the structure and regulation of cAMP-dependent protein kinase. *J. Mol. Biol.* 422, 215–229.
- Baum, G., Long, J. C., Jenkins, G. I., and Trewavas, A. J. (1999). Stimulation of the blue light phototropic receptor NPH1 causes a transient increase in cytosolic Ca<sup>2+</sup>. *Proc. Natl. Acad. Sci. U.S.A.* 96, 13554–13559.
- Benjamins, R., Ampudia, C. S., Hooykaas, P. J., and Offringa, R. (2003). PINOID-mediated signaling involves calcium-binding proteins. *Plant Physiol.* 132, 1623–1630.
- Benjamins, R., Quint, A., Weijers, D., Hooykaas, P., and Offringa, R. (2001). The PINOID protein kinase regulates organ development in *Arabidopsis* by enhancing polar auxin transport. *Development* 128, 4057–4067.
- Benschop, J. J., Mohammed, S., O’Flaherty, M., Heck, A. J., Sluiter, M., and Menke, F. L. (2007). Quantitative phosphoproteomics of early elicitor signaling in *Arabidopsis*. *Mol. Cell Proteomics* 6, 1198–1214.
- Biondi, R. M. (2004). Phosphoinositide-dependent protein kinase 1, a sensor of protein conformation. *Trends Biochem. Sci.* 29, 136–142.
- Biondi, R. M., Cheung, P. C., Casamayor, A., Deak, M., Currie, R. A., and Alessi, D. R. (2000). Identification of a pocket in the PDK1 kinase domain that interacts with PIF and the C-terminal residues of PKA. *EMBO J.* 19, 979–988.
- Biondi, R. M., Kieloch, A., Currie, R. A., Deak, M., and Alessi, D. R. (2001). The PIF-binding pocket in PDK1 is essential for activation of S6K and SGK, but not PKB. *EMBO J.* 20, 4380–4390.
- Blanc, G., Duncan, G., Agarkova, I., Borodovsky, M., Gurnon, J., Kuo, A., et al. (2010). The *Chlorella variabilis* NC64A genome reveals adaptation to photosymbiosis, coevolution with viruses, and cryptic sex. *Plant Cell* 22, 2943–2955.
- Bögner, L., Okresz, L., Henriques, R., and Anthony, R. G. (2003). Growth signalling pathways in *Arabidopsis* and the AGC protein kinases. *Trends Plant Sci.* 8, 424–431.
- Bouchard, R., Bailly, A., Blakeslee, J. J., Oehring, S. C., Vincenzetti, V., Lee, O. R., et al. (2006). Immunophilin-like TWISTED DWARF1 modulates auxin efflux activities of *Arabidopsis* P-glycoproteins. *J. Biol. Chem.* 281, 30603–30612.
- Camehl, I., Drzewiecki, C., Vadassery, J., Shahollari, B., Sherameti, I., Forzani, C., et al. (2011). The OX11 kinase pathway mediates *Piriformospora indica*-induced growth promotion in *Arabidopsis*. *PLoS Pathog.* 7, e1002051. doi:10.1371/journal.ppat.1002051
- Carafoli, E., Santella, L., Branca, D., and Brini, M. (2001). Generation, control, and processing of cellular calcium signals. *Crit. Rev. Biochem. Mol. Biol.* 36, 107–260.
- Casamayor, A., Morrice, N. A., and Alessi, D. R. (1999). Phosphorylation of Ser-241 is essential for the activity of 3-phosphoinositide-dependent protein kinase-1: identification of five sites of phosphorylation in vivo. *Biochem. J.* 342(Pt 2), 287–292.
- Chan, T. O., Rittenhouse, S. E., and Tschich, P. N. (1999). AKT/PKB and other D3 phosphoinositide-regulated kinases: kinase activation by phosphoinositide-dependent phosphorylation. *Annu. Rev. Biochem.* 68, 965–1014.
- Chen, Y., Hoehenwarter, W., and Weckwerth, W. (2010). Comparative analysis of phytohormone-responsive phosphoproteins in *Arabidopsis thaliana* using TiO<sub>2</sub>-phosphopeptide enrichment and mass accuracy precursor alignment. *Plant J.* 63, 1–17.
- Cheng, X., Ma, Y., Moore, M., Hemmings, B. A., and Taylor, S. S. (1998). Phosphorylation and activation of cAMP-dependent protein kinase by phosphoinositide-dependent protein kinase. *Proc. Natl. Acad. Sci. U.S.A.* 95, 9849–9854.
- Cheng, Y., Qin, G., Dai, X., and Zhao, Y. (2008). NPY genes and AGC kinases define two key steps in auxin-mediated organogenesis in *Arabidopsis*. *Proc. Natl. Acad. Sci. U.S.A.* 105, 21017–21022.
- Christensen, S. K., Dagenais, N., Chory, J., and Weigel, D. (2000). Regulation of auxin response by the protein kinase PINOID. *Cell* 100, 469–478.
- Christie, J. M. (2007). Phototropin blue-light receptors. *Annu. Rev. Plant Biol.* 58, 21–45.
- Christie, J. M., Yang, H., Richter, G. L., Sullivan, S., Thomson, C. E., Lin, J., et al. (2011). phot1 inhibition of ABCB19 primes lateral auxin fluxes in the shoot apex required for phototropism. *PLoS Biol.* 9, e1001076. doi:10.1371/journal.pbio.1001076
- Currie, R. A., Walker, K. S., Gray, A., Deak, M., Casamayor, A., Downes, C. P., et al. (1999). Role of phosphatidylinositol 3,4,5-trisphosphate in regulating the activity and localization of 3-phosphoinositide-dependent protein kinase-1. *Biochem. J.* 337(Pt 3), 575–583.
- Dai, M., Zhang, C., Kania, U., Chen, F., Xue, Q., McCray, T., et al. (2012). A PP6-type phosphatase holoenzyme directly regulates PIN phosphorylation and auxin efflux in *Arabidopsis*. *Plant Cell* 24, 2497–2514.



- Day, I. S., Miller, C., Golovkin, M., and Reddy, A. S. (2000). Interaction of a kinesin-like calmodulin-binding protein with a protein kinase. *J. Biol. Chem.* 275, 13737–13745.
- Deak, M., Casamayor, A., Currie, R. A., Downes, C. P., and Alessi, D. R. (1999). Characterisation of a plant 3-phosphoinositide-dependent protein kinase-1 homologue which contains a pleckstrin homology domain. *FEBS Lett.* 451, 220–226.
- Demarsy, E., Schepens, I., Okajima, K., Hersch, M., Bergmann, S., Christie, J., et al. (2012). Phytochrome Kinase Substrate 4 is phosphorylated by the phototropin 1 photoreceptor. *EMBO J.* 31, 3457–3467.
- Derelle, E., Ferraz, C., Rombauts, S., Rouze, P., Worden, A. Z., Robbens, S., et al. (2006). Genome analysis of the smallest free-living eukaryote *Ostreococcus tauri* unveils many unique features. *Proc. Natl. Acad. Sci. U.S.A.* 103, 11647–11652.
- Devarenne, T. P., Ekengren, S. K., Pedley, K. F., and Martin, G. B. (2006). Adi3 is a PDK1-interacting AGC kinase that negatively regulates plant cell death. *EMBO J.* 25, 255–265.
- Dhonukshe, P., Aniento, F., Hwang, I., Robinson, D. G., Mravec, J., Stierhof, Y. D., et al. (2007). Clathrin-mediated constitutive endocytosis of PIN auxin efflux carriers in *Arabidopsis*. *Curr. Biol.* 17, 520–527.
- Dhonukshe, P., Huang, F., Galvan-Ampudia, C. S., Mahonen, A. P., Kleine-Vehn, J., Xu, J., et al. (2010). Plasma membrane-bound AGC3 kinases phosphorylate PIN auxin carriers at TPRxS(N/S) motifs to direct apical PIN recycling. *Development* 137, 3245–3255.
- Ding, Z., Galvan-Ampudia, C. S., Demarsy, E., Langowski, L., Kleine-Vehn, J., Fan, Y., et al. (2011). Light-mediated polarization of the PIN3 auxin transporter for the phototropic response in *Arabidopsis*. *Nat. Cell Biol.* 13, 447–452.
- Dittrich, A. C., and Devarenne, T. P. (2012). Characterization of a PDK1 homologue from the moss *Physcomitrella patens*. *Plant Physiol.* 158, 1018–1033.
- Ek-Ramos, M. J., Avila, J., Cheng, C., Martin, G. B., and Devarenne, T. P. (2010). The T-loop extension of the tomato protein kinase AvrPto-dependent Pto-interacting protein 3 (Adi3) directs nuclear localization for suppression of plant cell death. *J. Biol. Chem.* 285, 17584–17594.
- Enugutti, B., Kirchhelle, C., Oelschner, M., Torres Ruiz, R. A., Schliebner, I., Leister, D., et al. (2012). Regulation of planar growth by the *Arabidopsis* AGC protein kinase UNICORN. *Proc. Natl. Acad. Sci. U.S.A.* 109, 15060–15065.
- Etchebehere, L. C., Van Bemmelen, M. X., Anjard, C., Traincard, F., Assemat, K., Reymond, C., et al. (1997). The catalytic subunit of *Dictyostelium* cAMP-dependent protein kinase – role of the N-terminal domain and of the C-terminal residues in catalytic activity and stability. *Eur. J. Biochem.* 248, 820–826.
- Folta, K. M., and Spalding, E. P. (2001). Unexpected roles for cryptochrome 2 and phototropin revealed by high-resolution analysis of blue light-mediated hypocotyl growth inhibition. *Plant J.* 26, 471–478.
- Friml, J., Yang, X., Michniewicz, M., Weijers, D., Quint, A., Tietz, O., et al. (2004). A PINOID-dependent binary switch in apical-basal PIN polar targeting directs auxin efflux. *Science* 306, 862–865.
- Frodin, M., Antal, T. L., Dummer, B. A., Jensen, C. J., Deak, M., Gammeltoft, S., et al. (2002). A phosphoserine/threonine-binding pocket in AGC kinases and PDK1 mediates activation by hydrophobic motif phosphorylation. *EMBO J.* 21, 5396–5407.
- Galvan-Ampudia, C. S., and Offringa, R. (2007). Plant evolution: AGC kinases tell the auxin tale. *Trends Plant Sci.* 12, 541–547.
- Gao, X., and Harris, T. K. (2006). Role of the PH domain in regulating in vitro autophosphorylation events required for reconstitution of PDK1 catalytic activity. *Bioorg. Chem.* 34, 200–223.
- Geldner, N., Anders, N., Wolters, H., Keicher, J., Kornberger, W., Muller, P., et al. (2003). The *Arabidopsis* GNOM ARF-GEF mediates endosomal recycling, auxin transport, and auxin-dependent plant growth. *Cell* 112, 219–230.
- Geldner, N., Friml, J., Stierhof, Y. D., Jürgens, G., and Palme, K. (2001). Auxin transport inhibitors block PIN1 cycling and vesicle trafficking. *Nature* 413, 425–428.
- Gleason, C., Chaudhuri, S., Yang, T., Munoz, A., Poovaiah, B. W., and Oldroyd, G. E. (2006). Nodulation independent of rhizobia induced by a calcium-activated kinase lacking autoinhibition. *Nature* 441, 1149–1152.
- Hammond, R. W., and Zhao, Y. (2000). Characterization of a tomato protein kinase gene induced by infection by *Potato spindle tuber viroid*. *Mol. Plant Microbe Interact.* 13, 903–910.
- Hanks, S. K., and Hunter, T. (1995). Protein kinases 6. The eukaryotic protein kinase superfamily: kinase (catalytic) domain structure and classification. *FASEB J.* 9, 576–596.
- Harada, A., Sakai, T., and Okada, K. (2003). Phot1 and phot2 mediate blue light-induced transient increases in cytosolic Ca<sup>2+</sup> differently in *Arabidopsis* leaves. *Proc. Natl. Acad. Sci. U.S.A.* 100, 8583–8588.
- Henrichs, S., Wang, B., Fukao, Y., Zhu, J., Charrier, L., Bailly, A., et al. (2012). Regulation of ABCB1/PGP1-catalysed auxin transport by linker phosphorylation. *EMBO J.* 31, 2965–2980.
- Hepler, P. K. (2005). Calcium: a central regulator of plant growth and development. *Plant Cell* 17, 2142–2155.
- Hrabak, E. M., Chan, C. W., Gribskov, M., Harper, J. F., Choi, J. H., Halford, N., et al. (2003). The *Arabidopsis* CDPK-SnRK superfamily of protein kinases. *Plant Physiol.* 132, 666–680.
- Huang, F., Zago, M. K., Abas, L., Van Marion, A., Galvan-Ampudia, C. S., and Offringa, R. (2010). Phosphorylation of conserved PIN motifs directs *Arabidopsis* PIN1 polarity and auxin transport. *Plant Cell* 22, 1129–1142.
- Huse, M., and Kuriyan, J. (2002). The conformational plasticity of protein kinases. *Cell* 109, 275–282.
- Inoue, S., Kinoshita, T., Matsumoto, M., Nakayama, K. I., Doi, M., and Shimazaki, K. (2008). Blue light-induced autophosphorylation of phototropin is a primary step for signaling. *Proc. Natl. Acad. Sci. U.S.A.* 105, 5626–5631.
- Inoue, S., Takemiya, A., and Shimazaki, K. (2010). Phototropin signaling and stomatal opening as a model case. *Curr. Opin. Plant Biol.* 13, 587–593.
- Isner, J. C., Nuhse, T., and Maathuis, F. J. (2012). The cyclic nucleotide cGMP is involved in plant hormone signalling and alters phosphorylation of *Arabidopsis thaliana* root proteins. *J. Exp. Bot.* 63, 3199–3205.
- Iwabuchi, K., Minamino, R., and Takagi, S. (2010). Actin reorganization underlies phototropin-dependent positioning of nuclei in *Arabidopsis* leaf cells. *Plant Physiol.* 152, 1309–1319.
- Jacobs, M., and Rubery, P. H. (1988). Naturally occurring auxin transport regulators. *Science* 241, 346–349.
- Jiang, T., and Qiu, Y. (2003). Interaction between Src and a C-terminal proline-rich motif of Akt is required for Akt activation. *J. Biol. Chem.* 278, 15789–15793.
- Kaiserli, E., Sullivan, S., Jones, M. A., Feeney, K. A., and Christie, J. M. (2009). Domain swapping to assess the mechanistic basis of *Arabidopsis* phototropin 1 receptor kinase activation and endocytosis by blue light. *Plant Cell* 21, 3226–3244.
- Kameshita, I., Shimomura, S., Nishio, K., Sueyoshi, N., Nishida, T., Nomura, M., et al. (2010). Expression and characterization of PKL01, an Ndr kinase homolog in *Lotus japonicus*. *J. Biochem.* 147, 799–807.
- Kannan, N., Haste, N., Taylor, S. S., and Neuwald, A. F. (2007). The hallmark of AGC kinase functional divergence is its C-terminal tail, a cis-acting regulatory module. *Proc. Natl. Acad. Sci. U.S.A.* 104, 1272–1277.
- Katayama, S., Sugiyama, Y., Hatano, N., Terachi, T., Sueyoshi, N., and Kameshita, I. (2012). PKL01, an NDR kinase homologue in plants, shows tyrosine kinase activity. *J. Biochem.* 152, 347–353.
- Kleine-Vehn, J., Huang, F., Naramoto, S., Zhang, J., Michniewicz, M., Offringa, R., et al. (2009). PIN auxin efflux carrier polarity is regulated by PINOID kinase-mediated recruitment into GNOM-independent trafficking in *Arabidopsis*. *Plant Cell* 21, 3839–3849.
- Kleine-Vehn, J., Langowski, L., Wisniewska, J., Dhonukshe, P., Brewer, P. B., and Friml, J. (2008). Cellular and molecular requirements for polar PIN targeting and transcytosis in plants. *Mol. Plant* 1, 1056–1066.
- Kleine-Vehn, J., Wabnick, K., Martinieri, A., Langowski, L., Willig, K., Naramoto, S., et al. (2011). Recycling, clustering, and endocytosis jointly maintain PIN auxin carrier polarity at the plasma membrane. *Mol. Syst. Biol.* 7, 540.
- Kong, S. G., Suzuki, T., Tamura, K., Mochizuki, N., Hara-Nishimura, I., and Nagatani, A. (2006). Blue light-induced association of phototropin 2 with the Golgi apparatus. *Plant J.* 45, 994–1005.
- Lariguet, P., and Dunand, C. (2005). Plant photoreceptors: phylogenetic overview. *J. Mol. Evol.* 61, 559–569.
- Lariguet, P., Schepens, I., Hodgson, D., Pedmale, U. V., Trevisan, M., Kami, C., et al. (2006). PHYTOCHROME KINASE SUBSTRATE 1 is a phototropin 1 binding protein required for phototropism. *Proc. Natl. Acad. Sci. U.S.A.* 103, 10134–10139.
- Lewis, D. R., Ramirez, M. V., Miller, N. D., Vallabhaneni, P., Ray, W. K., Helm, R. F., et al. (2011). Auxin and ethylene induce flavonol accumulation through distinct transcriptional networks. *Plant Physiol.* 156, 144–164.

- Li, H., Lin, D., Dhonukshe, P., Nagawa, S., Chen, D., Friml, J., et al. (2011). Phosphorylation switch modulates the interdigitated pattern of PIN1 localization and cell expansion in *Arabidopsis* leaf epidermis. *Cell Res.* 21, 970–978.
- Mahfouz, M. M., Kim, S., Delauney, A. J., and Verma, D. P. (2006). *Arabidopsis* TARGET OF RAPAMYCIN interacts with RAPTOR, which regulates the activity of S6 kinase in response to osmotic stress signals. *Plant Cell* 18, 477–490.
- Manning, G., Plowman, G. D., Hunter, T., and Sudarsanam, S. (2002). Evolution of protein kinase signaling from yeast to man. *Trends Biochem. Sci.* 27, 514–520.
- Masters, T. A., Calleja, V., Armoogum, D. A., Marsh, R. J., Applebee, C. J., Laguerre, M., et al. (2010). Regulation of 3-phosphoinositide-dependent protein kinase 1 activity by homodimerization in live cells. *Sci. Signal.* 3, ra78.
- Matsui, H., Miyao, A., Takahashi, A., and Hirochika, H. (2010). PDK1 kinase regulates basal disease resistance through the OsOx1l-OsPti1a phosphorylation cascade in rice. *Plant Cell Physiol.* 51, 2082–2091.
- McLoughlin, E., Galvan-Ampudia, C. S., Julkowska, M. M., Caarls, L., Van Der Does, D., Lauriere, C., et al. (2012). The Snf1-related protein kinases SnRK2.4 and SnRK2.10 are involved in maintenance of root system architecture during salt stress. *Plant J.* 72, 436–449.
- McSteen, P., Malcomber, S., Skirpan, A., Lunde, C., Wu, X., Kellogg, E., et al. (2007). *barren inflorescence2* encodes a co-ortholog of the PINOID serine/threonine kinase and is required for organogenesis during inflorescence and vegetative development in maize. *Plant Physiol.* 144, 1000–1011.
- Men, S., Boutte, Y., Ikeda, Y., Li, X., Palme, K., Stierhof, Y. D., et al. (2008). Sterol-dependent endocytosis mediates post-cytokinetic acquisition of PIN2 auxin efflux carrier polarity. *Nat. Cell Biol.* 10, 237–244.
- Michniewicz, M., Zago, M. K., Abas, L., Weijers, D., Schweighofer, A., Meskiene, I., et al. (2007). Antagonistic regulation of PIN phosphorylation by PP2A and PINOID directs auxin flux. *Cell* 130, 1044–1056.
- Miranda-Saavedra, D., and Barton, G. J. (2007). Classification and functional annotation of eukaryotic protein kinases. *Proteins* 68, 893–914.
- Mora, A., Komander, D., Van Aalten, D. M., and Alessi, D. R. (2004). PDK1, the master regulator of AGC kinase signal transduction. *Semin. Cell Dev. Biol.* 15, 161–170.
- Morita, Y., and Kyoizuka, J. (2007). Characterization of OsPID, the rice ortholog of PINOID, and its possible involvement in the control of polar auxin transport. *Plant Cell Physiol.* 48, 540–549.
- Mravec, J., Kubes, M., Bielach, A., Gaykova, V., Petrasko, J., Skupa, P., et al. (2008). Interaction of PIN and PGP transport mechanisms in auxin distribution-dependent development. *Development* 135, 3345–3354.
- Nagawa, S., Xu, T., Lin, D., Dhonukshe, P., Zhang, X., Friml, J., et al. (2012). ROP GTPase-dependent actin microfilaments promote pin1 polarization by localized inhibition of clathrin-dependent endocytosis. *PLoS Biol.* 10, e1001299. doi:10.1371/journal.pbio.1001299
- Newton, A. C. (2001). Protein kinase C: structural and spatial regulation by phosphorylation, cofactors, and macromolecular interactions. *Chem. Rev.* 101, 2353–2364.
- Nirula, A., Ho, M., Phee, H., Roose, J., and Weiss, A. (2006). Phosphoinositide-dependent kinase 1 targets protein kinase A in a pathway that regulates interleukin 4. *J. Exp. Med.* 203, 1733–1744.
- Nolen, B., Taylor, S., and Ghosh, G. (2004). Regulation of protein kinases; controlling activity through activation segment conformation. *Mol. Cell* 15, 661–675.
- Nuhse, T. S., Stensballe, A., Jensen, O. N., and Peck, S. C. (2004). Phosphoproteomics of the *Arabidopsis* plasma membrane and a new phosphorylation site database. *Plant Cell* 16, 2394–2405.
- Onodera, A., Kong, S. G., Doi, M., Shimazaki, K., Christie, J., Mochizuki, N., et al. (2005). Phototropin from *Chlamydomonas reinhardtii* is functional in *Arabidopsis thaliana*. *Plant Cell Physiol.* 46, 367–374.
- Otterhag, L., Gustavsson, N., Alsterfjord, M., Pical, C., Lehrach, H., Gobom, J., et al. (2006). *Arabidopsis* PDK1: identification of sites important for activity and downstream phosphorylation of S6 kinase. *Biochimie* 88, 11–21.
- Oyama, T., Shimura, Y., and Okada, K. (2002). The *IRE* gene encodes a protein kinase homologue and modulates root hair growth in *Arabidopsis*. *Plant J.* 30, 289–299.
- Pearce, L. R., Komander, D., and Alessi, D. R. (2010). The nuts and bolts of AGC protein kinases. *Nat. Rev. Mol. Cell Biol.* 11, 9–22.
- Pedmale, U. V., and Liscum, E. (2007). Regulation of phototropic signaling in *Arabidopsis* via phosphorylation state changes in the phototropin 1-interacting protein NPH3. *J. Biol. Chem.* 282, 19992–20001.
- Petersen, L. N., Ingle, R. A., Knight, M. R., and Denby, K. J. (2009). OXII protein kinase is required for plant immunity against *Pseudomonas syringae* in *Arabidopsis*. *J. Exp. Bot.* 60, 3727–3735.
- Pfeifer, A., Mathes, T., Lu, Y., Hegemann, P., and Kottke, T. (2010). Blue light induces global and localized conformational changes in the kinase domain of full-length phototropin. *Biochemistry* 49, 1024–1032.
- Pislaru, C. I., and Dickstein, R. (2007). An IRE-like AGC kinase gene, *MtIRE*, has unique expression in the invasion zone of developing root nodules in *Medicago truncatula*. *Plant Physiol.* 144, 682–694.
- Reitel, M. C., Lecourieux, D., Ouaked, F., Usher, S. L., Petersen, L., Okamoto, H., et al. (2004). OXII kinase is necessary for oxidative burst-mediated signalling in *Arabidopsis*. *Nature* 427, 858–861.
- Roberts, D., Pedmale, U. V., Morrow, J., Sachdev, S., Lechner, E., Tang, X., et al. (2011). Modulation of phototropic responsiveness in *Arabidopsis* through ubiquitination of phototropin 1 by the CUL3-Ring E3 ubiquitin ligase CRL3(NPH3). *Plant Cell* 23, 3627–3640.
- Sakamoto, K., and Briggs, W. R. (2002). Cellular and subcellular localization of phototropin 1. *Plant Cell* 14, 1723–1735.
- Salomon, M., Christie, J. M., Knieb, E., Lempert, U., and Briggs, W. R. (2000). Photochemical and mutational analysis of the FMN-binding domains of the plant blue light receptor, phototropin. *Biochemistry* 39, 9401–9410.
- Salomon, M., Knieb, E., Von Zepelin, T., and Rudiger, W. (2003). Mapping of low- and high-fluence autophosphorylation sites in phototropin 1. *Biochemistry* 42, 4217–4225.
- Santner, A. A., and Watson, J. C. (2006). The WAG1 and WAG2 protein kinases negatively regulate root waving in *Arabidopsis*. *Plant J.* 45, 752–764.
- Sastri, M., Barraclough, D. M., Carmichael, P. T., and Taylor, S. S. (2005). A kinase-interacting protein localizes protein kinase A in the nucleus. *Proc. Natl. Acad. Sci. U.S.A.* 102, 349–354.
- Sathyanarayanan, P. V., Cremo, C. R., and Poovaiah, B. W. (2000). Plant chimeric Ca<sup>2+</sup>-Calmodulin-dependent protein kinase. Role of the neural visinin-like domain in regulating autophosphorylation and calmodulin affinity. *J. Biol. Chem.* 275, 30417–30422.
- Sieburth, L. E., Muday, G. K., King, E. J., Benton, G., Kim, S., Metcalf, K. E., et al. (2006). SCARFACE encodes an ARF-GAP that is required for normal auxin efflux and vein patterning in *Arabidopsis*. *Plant Cell* 18, 1396–1411.
- Skirpan, A., Culler, A. H., Gallavotti, A., Jackson, D., Cohen, J. D., and McSteen, P. (2009). BARREN INFLORESCENCE2 interaction with ZmPIN1a suggests a role in auxin transport during maize inflorescence development. *Plant Cell Physiol.* 50, 652–657.
- Skirpan, A., Wu, X., and McSteen, P. (2008). Genetic and physical interaction suggest that BARREN STALK 1 is a target of BARREN INFLORESCENCE2 in maize inflorescence development. *Plant J.* 55, 787–797.
- Sukumar, P., Edwards, K. S., Rahman, A., Delong, A., and Muday, G. K. (2009). PINOID kinase regulates root gravitropism through modulation of PIN2-dependent basipetal auxin transport in *Arabidopsis*. *Plant Physiol.* 150, 722–735.
- Sullivan, S., Thomson, C. E., Kaiserli, E., and Christie, J. M. (2009). Interaction specificity of *Arabidopsis* 14-3-3 proteins with phototropin receptor kinases. *FEBS Lett.* 583, 2187–2193.
- Swainsbury, D. J., Zhou, L., Oldroyd, G. E., and Bornemann, S. (2012). Calcium ion binding properties of *Medicago truncatula* calcium/calmodulin-dependent protein kinase. *Biochemistry* 51, 6895–6907.
- Swartz, T. E., Corchano, S. B., Christie, J. M., Lewis, J. W., Szundi, I., Briggs, W. R., et al. (2001). The photocycle of a flavin-binding domain of the blue light photoreceptor phototropin. *J. Biol. Chem.* 276, 36493–36500.
- Takayama, Y., Nakasako, M., Okajima, K., Iwata, A., Kashojiya, S., Matsui, Y., et al. (2011). Light-induced movement of the LOV2 domain in an Asp720Asn mutant LOV2-kinase fragment of *Arabidopsis* phototropin 2. *Biochemistry* 50, 1174–1183.
- Taylor, S. S., and Kornev, A. P. (2011). Protein kinases: evolution of dynamic regulatory proteins. *Trends Biochem. Sci.* 36, 65–77.
- Tholey, A., Pipkorn, R., Bossemeyer, D., Kinzel, V., and Reed, J. (2001).

- Influence of myristoylation, phosphorylation, and deamidation on the structural behavior of the N-terminus of the catalytic subunit of cAMP-dependent protein kinase. *Biochemistry* 40, 225–231.
- Tseng, T. S., Whipps, C., Hangarter, R. P., and Briggs, W. R. (2012). The role of a 14-3-3 protein in stomatal opening mediated by PHOT2 in *Arabidopsis*. *Plant Cell* 24, 1114–1126.
- Wick, M. J., Ramos, F. J., Chen, H., Quon, M. J., Dong, L. Q., and Liu, F. (2003). Mouse 3-phosphoinositide-dependent protein kinase-1 undergoes dimerization and trans-phosphorylation in the activation loop. *J. Biol. Chem.* 278, 42913–42919.
- Xiong, Y., and Sheen, J. (2012). Rapamycin and glucose-target of rapamycin (TOR) protein signaling in plants. *J. Biol. Chem.* 287, 2836–2842.
- Xu, J., and Scheres, B. (2005). Dissection of *Arabidopsis* ADP-RIBOSYLATION FACTOR 1 function in epidermal cell polarity. *Plant Cell* 17, 525–536.
- Xu, T., Wen, M., Nagawa, S., Fu, Y., Chen, J. G., Wu, M. J., et al. (2010). Cell surface- and rho GTPase-based auxin signaling controls cellular interdigitation in *Arabidopsis*. *Cell* 143, 99–110.
- Yonemoto, W., McGlone, M. L., Grant, B., and Taylor, S. S. (1997). Autophosphorylation of the catalytic subunit of cAMP-dependent protein kinase in *Escherichia coli*. *Protein Eng.* 10, 915–925.
- Zegzouti, H., Anthony, R. G., Jahchan, N., Bögre, L., and Christensen, S. K. (2006a). Phosphorylation and activation of PINOID by the phospholipid signaling kinase 3-phosphoinositide-dependent protein kinase 1 (PDK1) in *Arabidopsis*. *Proc. Natl. Acad. Sci. U.S.A.* 103, 6404–6409.
- Zegzouti, H., Li, W., Lorenz, T. C., Xie, M., Payne, C. T., Smith, K., et al. (2006b). Structural and functional insights into the regulation of *Arabidopsis* AGC VIIa kinases. *J. Biol. Chem.* 281, 35520–35530.
- Zhang, J., Nodzynski, T., Pencik, A., Rolcik, J., and Friml, J. (2010). PIN phosphorylation is sufficient to mediate PIN polarity and direct auxin transport. *Proc. Natl. Acad. Sci. U.S.A.* 107, 918–922.
- Zhang, Y., He, J., and McCormick, S. (2009). Two *Arabidopsis* AGC kinases are critical for the polarized growth of pollen tubes. *Plant J.* 58, 474–484.
- Zhang, Y., and McCormick, S. (2009). AGCVIII kinases: at the crossroads of cellular signaling. *Trends Plant Sci.* 14, 689–695.
- Zourelidou, M., Muller, I., Willige, B. C., Nill, C., Jikumaru, Y., Li, H., et al. (2009). The polarly localized D6 PROTEIN KINASE is required for efficient auxin transport in *Arabidopsis thaliana*. *Development* 136, 627–636.

**Conflict of Interest Statement:** The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Received: 13 July 2012; accepted: 22 October 2012; published online: 16 November 2012.

Citation: Rademacher EH and Offringa R (2012) Evolutionary adaptations of plant AGC kinases: from light signaling to cell polarity regulation. *Front. Plant Sci.* 3:250. doi: 10.3389/fpls.2012.00250

This article was submitted to *Frontiers in Plant Traffic and Transport*, a specialty of *Frontiers in Plant Science*.

Copyright © 2012 Rademacher and Offringa. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits use, distribution and reproduction in other forums, provided the original authors and source are credited and subject to any copyright notices concerning any third-party graphics etc.